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(54) Title: DETECTION OF NEGATIVE-STRAND RNA VIRUSES (57) Abstract A diagnostic assay for detecting a negative-strand RNA virus in a sample and a genetically engineered cell for use in the assay are disclosed. The cell expresses a heterologous DNA-dependent RNA polymerase that synthesizes a minigenome or miniantigenome of the RNA virus from a cDNA template present in the cell. The cell also expresses the nucleocapsid proteins of the negative-strand virus that are necessary for replication of the minigenome or miniantigenome. Infection of the cell by the negative-strand virus results in expression of a reporter gene product encoded by the miniantigenome.		

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DETECTION OF NEGATIVE-STRAND RNA VIRUSES

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Background of the Invention

(1) Field of the Invention

This invention generally relates to the field of virology and, more particularly, to a method for detecting a negative-strand RNA virus in a biological specimen and genetically engineered cells for use in the method.

(2) Description of Related Art

Numerous negative-strand RNA viruses are pathogenic to humans and other animals. Examples of human diseases caused by negative-strand viruses include mumps, measles, pneumonia, bronchitis, influenza, infectious croup, rabies, ebola hemorrhagic fever, marburg hemorrhagic fever, and LaCrosse encephalitis. Thus,

detection of negative-strand viruses in biological specimens is important clinically and for various research purposes.

Although there is considerable diversity in the genomic structure and biological properties of negative-strand RNA viruses, the RNA replication and transcription strategies of these viruses have common features. As with all RNA viruses, negative-strand RNA viruses express an RNA-dependent RNA polymerase and other RNA replicase and transcriptase factors necessary to transcribe their mRNA and replicate their genomes (Olivo, P.D., *Clin. Microb. Rev.* 9:321-334, 1996). In the virion of a negative-strand virus, the RNA polymerase forms a complex with the nucleocapsid protein and the genomic RNA. Once the virion enters the cell and begins to uncoat, the virion RNA polymerase transcribes subgenomic mRNAs from the genomic RNA. The RNA polymerase also synthesizes full-length positive-strand replicative-intermediate RNA (antigenome), which is used as a template for making many copies of negative-stranded genomic RNA that then are used as templates for secondary transcription of additional viral mRNAs.

The study of negative-strand viruses has been complicated by the fact that the RNA-dependent RNA polymerase uses as template only RNA associated with virus-specific nucleocapsid proteins; thus, naked genomic RNA transfected into a cell is not replicated (Bukreyev et al., *J. Virol.* 70:6634-6641, 1996; Olivo, *supra*). Recently, however, a number of laboratories have described advances in the genetic manipulation of various negative-strand RNA viruses. For example, infectious rabies virus, Sendai virus, vesicular stomatitis virus, measles virus, and respiratory syncytial virus (RSV) have been reportedly recovered by using T7 RNA polymerase to generate a full-length antigenomic transcript from a cDNA of the genome in the cell cytoplasm together with the viral proteins necessary for assembly of a nucleocapsid and for RNA replication and transcription (Schnell et al., *EMBO J* 13:4195-4203, 1994; Garcin et al., *EMBO J* 14:6087-6094, 1995; Lawson et al., *Proc. Natl. Acad. Sci. USA* 92:4477-4481, 1995; Radecke et al., *EMBO J* 14:5773-5784, 1995; Whelan et al., *Proc. Natl. Acad. Sci. USA* 92:8388-8392, 1995; Collins et al., *Proc. Natl. Acad. Sci. USA* 92:11563-11567, 1995). In these reports, the T7 RNA polymerase was provided to the cell cytoplasm by infection with a recombinant vaccinia virus containing the gene for T7 RNA polymerase.

Other studies investigating viral protein function in replication and transcription of negative-strand RNA viruses have employed "minigenomes", in which some or all of the viral protein-coding sequences are replaced with a reporter gene flanked by *cis*-acting elements necessary for replication and transcription. See, 5 e.g., Grosfeld et al., *J. of Virol.* 69:5677-5686, 1995; Conzelmann et al., *J. Virol.* 68:713-719, 1994; De et al., *Virol.* 196:344-348, 1993; Dimock et al., *J. Virol.* 67:2772-2778, 1993. In this approach, a cDNA is constructed in which the minigenome is operably linked to a promoter for a bacteriophage RNA polymerase, and the minigenome cDNA is transfected into cells and transcribed by the 10 bacteriophage RNA polymerase in the presence of various viral proteins supplied by cotransfected plasmids, whose expression is also driven by the bacteriophage RNA polymerase, which is supplied by infection with a recombinant vaccinia virus. Induction of reporter gene expression indicates the transfected cell is expressing all the viral proteins needed to replicate and transcribe the minigenome.

15 Using this approach to study replication and transcription in RSV, the major nucleocapsid protein (N), the nucleocapsid-associated phosphoprotein (P), and the polymerase L protein were identified as sufficient to replicate the minigenome (Grosfeld et al., *supra*; Yu et al., *J. Virol.* 69:2412-2419, 1995). Coexpression of the N, P, and L proteins with a minigenome containing the *cat* reporter gene also resulted 20 in synthesis of full-size and incomplete CAT mRNA species and abundant expression of CAT (Grosfeld et al., *supra*). In contrast, efficient synthesis of full-length mRNA was observed when cells containing the minigenome cDNA and the N, P and L expression plasmids were coinfectd with the T7-expressing vaccinia virus and RSV, i.e., the minigenome RNA and the N, P and L proteins were synthesized by T7 RNA 25 polymerase in the presence of infectious RSV (Grosfeld et al., *supra*). A later study demonstrated that expression of an additional RSV protein, the transcription elongation factor M2-1, was required for fully processive sequential transcription in this T7-expression plasmid complementation system (Collins et al., *Proc. Natl. Acad. Sci. USA* 93:81-85, 1996).

30 In U.S. Patent No. 5,591,579, a genetically engineered cell line and method for detecting positive-strand RNA viruses was described. The cell line is stably transformed with a cDNA copy of a structurally defective RNA virus genome which contains (1) the *cis*-acting sequences of the RNA virus genome which are necessary

for replication and transcription by trans-acting enzymes from the RNA virus and (2) a reporter gene. The cDNA is constitutively transcribed into a (+) strand RNA molecule from a RNA polymerase II promoter in the nucleus of the host cell, but little or no expression of the reporter gene occurs until the cell is infected by a positive-strand RNA virus that recognizes the *cis*-acting sequences and causes significant replication of the (+) strand RNA molecule through a (-) strand RNA intermediate.

Recently, a review article on the use of transgenic cell lines for detecting animal viruses speculated that a similar strategy could be used to make a cell line for detecting negative-strand RNA viruses (Olivo et al., *supra*). The article stated generally that such a cell line would constitutively express the viral nucleocapsid protein and a chimeric antigenomic RNA molecule which contains a reporter gene open reading frame (ORF) and the *cis*-acting sequences necessary for replication and transcription by the replicase and transcriptase of the same negative-strand RNA virus. This chimeric RNA molecule would be designed, in an unspecified manner, to preclude translation of the reporter gene in uninfected cells. Replication and transcription of the chimeric RNA molecule and synthesis of translatable reporter gene mRNA would be carried out by the replicase-transcriptase complex brought into this hypothetical cell by infection with the negative-strand RNA virus. However, this article did not teach how the nucleocapsid protein and chimeric RNA molecule would be constitutively expressed or how to design the chimeric RNA molecule to preclude translation of the reporter gene in the absence of infectious virus. It would be desirable, therefore, to provide a rapid, specific, sensitive and cost-efficient assay for the detection of infectious negative-stranded RNA viruses.

Summary of the Invention

Briefly, therefore, the present invention is directed to novel compositions and methods which permit the detection of a negative-strand RNA virus in a specimen. In one embodiment, the invention provides a genetically engineered cell which expresses a reporter gene product only in the presence of infectious virions of the negative-strand RNA virus. The genetically engineered cell comprises a polynucleotide encoding a DNA-dependent RNA polymerase and a cDNA comprising a minigenome and a miniantigenome of the negative-strand RNA virus operably linked to a promoter for the DNA-dependent RNA polymerase. The miniantigenome comprises a

nucleotide sequence encoding a reporter gene product whose expression is dependent upon the presence of the negative-strand RNA virus.

Expression of the DNA-dependent RNA polymerase in the cell results in transcription of the cDNA into a minigenome RNA or an miniantigenome RNA, depending on the orientation of the promoter with respect to the open reading frame (ORF) of the reporter gene. When the promoter is located downstream of the ORF, a minigenome RNA is synthesized which contains an untranslatable negative-sense copy of the reporter gene ORF and when the promoter is located upstream of the ORF, a miniantigenome RNA is synthesized. Although this miniantigenome contains a positive-sense copy of the reporter gene ORF, its translation is very inefficient because the miniantigenome is not a capped, polyadenylated mRNA. Thus, detectable expression of the reporter gene in the absence of infectious virus does not occur. The cell also comprises one or more nucleotide sequences encoding each of the nucleocapsid proteins of the negative-strand RNA virus which are necessary and sufficient for replication of the minigenome RNA or miniantigenome RNA synthesized by the DNA-dependent RNA polymerase. Minigenome RNA synthesized in the presence of these nucleocapsid proteins, which includes minigenome RNA made as an intermediate during replication of the miniantigenome, is stabilized and amplified without detectable expression of the reporter gene product. Infection of the cell with the negative-strand virus introduces all the viral gene products necessary for transcription of the preformed minigenome RNA to produce translatable mRNA molecules encoding the reporter gene product.

In another embodiment, the invention provides a method for detecting a negative-strand RNA virus in a sample which comprises culturing the above-described genetically engineered cell for a time sufficient to synthesize the minigenome or miniantigenome RNA and the nucleocapsid proteins, then incubating the cells with the sample and detecting expression of the reporter gene product. In some embodiments, the method includes quantifying the number of infectious particles of the RNA virus that are present in the sample.

In other embodiments the genetically engineered cell is used to screen compounds for anti-viral activity and for measuring antibody responses of individuals infected with or vaccinated against the RNA virus.

In yet another embodiment, the invention provides a kit for detecting the presence of a negative-strand RNA virus in a sample which comprises a supply of the above-described genetically engineered cells. In a preferred embodiment, the kit also includes a set of reagents for detecting expression of the reporter gene product.

5 The invention also provides a method for making a genetically-engineered cell capable of detecting a negative-strand RNA virus. The method comprises (1) providing a cell that is susceptible to infection by the negative-strand RNA virus; (2) introducing into the cell a polynucleotide encoding a DNA-dependent RNA polymerase and selecting for cells containing the polynucleotide; (3) introducing into
10 the selected cells a cDNA comprising a minigenome and an miniantigenome of the negative-strand RNA virus operably linked to a promoter for the DNA-dependent RNA polymerase, wherein the miniantigenome comprises a nucleotide sequence encoding a reporter gene product, and wherein expression of the reporter gene product is dependent upon the presence of the negative-strand RNA virus; and (3) introducing
15 into the cells one or more nucleotide sequences encoding each of the nucleocapsid proteins of the negative-strand RNA virus which are necessary and sufficient for replication of minigenome RNA or miniantigenome RNA synthesized by the DNA-dependent RNA polymerase.

 Among the several advantages of the present invention may be noted the
20 provision of a genetically engineered cell which expresses a reporter gene product only after infection by a particular negative-strand RNA virus; the provision of a rapid and sensitive assay for specifically detecting the negative-strand RNA virus; the provision of such an assay that is capable of quantitating the amount of the negative-strand RNA virus in a sample that can be useful for assessing compounds for antiviral
25 activity and for measuring antibody responses in infected or vaccinated individuals; and the provision of a kit for detecting and/or quantitating the negative-strand RNA virus.

Brief Description of the Drawings

30 Figure 1A is a schematic representation of SR19/pac cDNA, showing an SP6 promoter operably linked to a DNA copy of a noncytopathic Sindbis virus replicon in which the gene for puromycin acetyltransferase (PAC) is operably linked to the Sindbis subgenomic promoter;

Figure 1B is a schematic representation of SR19/T7pol/pac cDNA, which is identical to the cDNA of Figure 1A except that the noncytopathic Sindbis virus replicon contains a the T7 RNA polymerase operably linked to a second Sindbis subgenomic promoter;

Figure 2 is a bar graph of β -galactosidase activity in baby hamster kidney (BHK) cells transformed with a noncytopathic Sindbis virus replicon expressing T7 RNA polymerase (SR19/T7/pac) or the parent replicon (SR19/pac) and transfected with a T7 expression plasmid expressing the *E. coli lacZ* gene under the control of the T7 promoter (pTM1-LacZ) or the parent T7 expression plasmid (pTM1);

Figure 3A is a bar graph of β -galactosidase activity in mock-infected or RSV-infected SR19/T7/pac-BHK or SR19/pac-BHK cells transfected with a cDNA of a *lacZ*-containing RSV minigenome (pMP210LacZ) alone or with T7 expression plasmids expressing the RSV nucleocapsid proteins N, L and P (+ NLP);

Figure 3B is a bar graph of β -galactosidase activity in mock-infected or RSV-infected SR19/T7/pac-BHK or SR19/pac-BHK cells transfected with pMP210LacZ alone or with various combinations of the T7 expression plasmids for N, L and P as indicated by the + and - symbols below the graph;

Figure 4A is a graph of β -galactosidase activity in SR19/T7/pac-BHK cells cotransfected with pMP210LacZ and the nucleocapsid expression plasmids and infected with the indicated amounts of RSV;

Figure 4B is a graph of β -galactosidase activity in the cells from Figure 4A at the indicated times following RSV infection at a high multiplicity of infection (HMIO) or at a low multiplicity of infection (LMOI);

Figure 5 illustrates the effect of ribavarin on the induction of reporter gene activity in RSV-infected SR19/T7/pac-BHK cells cotransfected with pMP210LacZ and the nucleocapsid expression plasmids showing in FIG. 5A a bar graph of β -galactosidase activity at high concentrations of ribavarin and in FIG. 5B a dose-response curve of β -galactosidase activity at subinhibitory concentrations or ribavarin expressed as a percentage of the control;

Figure 6 is a graph of β -galactosidase activity in SR19/T7/pac-BHK cells cotransfected with pMP210LacZ and the nucleocapsid expression plasmids after infection with RSV which had been preincubated with the indicated dilutions of pooled human immune globulin; and

Figure 7 is a schematic representation of the recombinant constructs and molecular pathways involved in generating a reporter gene product in accordance with one embodiment of the invention.

5 Description of Preferred Embodiments

In accordance with the present invention, a method for detecting negative-strand RNA viruses and genetically engineered cells for use in the method are provided. The invention is based on the inventors' discoveries (1) that minigenome RNA of a negative-strand virus expressed by a genetically engineered cell in the
10 presence of certain viral nucleocapsid proteins is accessible for transcription of translatable reporter gene mRNA when the cell is subsequently infected with the negative-strand virus and (2) that expression of the product of the reporter gene is not detectably expressed in the absence of the negative-strand virus.

In the context of this disclosure, the following terms shall be defined as
15 follows unless otherwise indicated:

"cis-acting sequences" means the nucleotide sequences from an RNA virus genome that are necessary for the recognition of the genomic RNA by specific protein(s) of the RNA virus that carry out replication and/or transcription of the genome;

20 "heterologous DNA-dependent RNA polymerase" means a DNA-dependent RNA polymerase not naturally encoded in a vertebrate cell;

"infectious" when used to describe a virus or an RNA molecule, means a virus or RNA molecule that is self-replicating and provides for transcription of viral mRNAs in a host cell;

25 "miniantigenome" means an RNA molecule or a DNA molecule that is complementary to a minigenome;

"minigenome" means a DNA or RNA analog of the genome of a negative-strand RNA virus which contains the cis-acting sequences necessary for replication and transcription in cells infected with the negative-strand RNA virus and which
30 contains a negative-sense open reading frame of a reporter gene in place of one or more of the viral protein-coding sequences;

"negative-strand RNA virus" means a virus whose genome consists of one negative-sense RNA molecule (nonsegmented genome) or a plurality of RNA

molecules of negative sense (segmented genome) and which replicates its genome through a positive-sense RNA intermediate(s);

"replicon" means a replication-competent viral RNA that contains the genetic information needed for virus replication but not for virus assembly;

5 "transfected cell" or "transformed cell" means a cell containing an exogenously introduced nucleic acid molecule which may be present in the nucleus or the cytoplasm of the cell;

"stably transformed cell" means a cell containing an exogenously introduced nucleic acid molecule which is present in the nucleus of the cell and may be stably
10 integrated into the chromosomal DNA of the cell.

A genetically engineered cell according to the present invention is a vertebrate cell that is susceptible to infection by the negative-strand RNA virus to be detected and that is capable of expressing a heterologous DNA-dependent RNA polymerase which synthesizes a minigenome RNA or miniantigenome RNA of a negative-strand
15 virus from a promoter recognized by the DNA-dependent RNA polymerase. The cell is also capable of expressing the nucleocapsid proteins of the negative-strand virus that are necessary for replication of the minigenome RNA or miniantigenome RNA.

A cell is susceptible to infection if the infecting virus can enter the cell and proceed far enough in its replication cycle to express proteins necessary for efficient
20 synthesis of translatable reporter gene mRNA from preformed minigenome RNA. It will be understood by those skilled in the art that cell lines susceptible to infection by a particular negative-strand RNA virus can be readily identified by searching the literature for known susceptible cell lines and/or by screening candidate cell lines using well-known procedures requiring only routine experimentation. Susceptible
25 cells for negative-strand RNA viruses include, but are not limited to baby hamster kidney cells, African green monkey cells, rabbit skin fibroblasts, 3T3 mouse cells and the like. Baby hamster kidney cells are preferred for use as the host cell for detecting negative-strand viruses that infect mammals, including humans.

The heterologous DNA-dependent RNA polymerase expressed by the cell comprises
30 a naturally-occurring amino acid sequence or a functional equivalent thereof. A functional equivalent of a DNA-dependent RNA polymerase contains one or more amino acid substitutions, additions or deletions in the naturally-occurring amino acid sequence that do not prevent the modified polymerase from synthesizing full-length transcripts from a DNA

template. Typically, the heterologous DNA-dependent RNA polymerase comprises the amino acid sequence of an RNA polymerase made by a bacteriophage such as T7, T3 or SP6. A preferred DNA-dependent RNA polymerase for use in the present invention is T7 RNA polymerase.

5 In one embodiment, the heterologous DNA-dependent RNA polymerase is encoded by a polynucleotide integrated into the cell nucleus and comprises a polymerase II promoter recognized by the host cell transcription machinery operably linked to a nucleotide sequence encoding the heterologous DNA-dependent RNA polymerase. In a preferred embodiment, the polynucleotide encoding the

10 heterologous DNA-dependent RNA polymerase is an RNA molecule that comprises a noncytopathic positive-strand virus replicon. Use of a noncytopathic replicon allows for long-term, continuous expression of the DNA-dependent RNA polymerase without killing the host cell. Modification of replicons to make them noncytopathic is known in the art and can be accomplished, i.e., by isolating variants having adaptive

15 mutations in the viral replication machinery that allows persistent noncytopathic replication in vertebrate cells or by using a dominant selectable marker (see e.g., Frolov et al., *Proc. Natl. Acad. Sci. USA* 93:11371-11377, 1996 and referenced cited therein). Preferred viruses for preparation of noncytopathic replicons include flaviviruses, e.g., Kunjin virus, alphaviruses such as Sindbis virus, Semliki forest

20 virus, nodaviruses such as flock house virus, astroviruses and coronaviruses. The noncytopathic replicon can be transcribed from a cDNA of the replicon that is operably linked to a polymerase II promoter and integrated into the nucleus. Alternatively, the replicon can be prepared chemically or by *in vitro* transcription from a plasmid and then transfected into the cell cytoplasm.

25 The heterologous DNA-dependent RNA polymerase specifically recognizes a promoter in a cDNA comprising a minigenome and a miniantigenome of the negative-strand RNA virus to be detected. Typically, the nucleotide sequence of the promoter is identical to a promoter sequence found in the same organism that encodes the DNA-dependent RNA polymerase. However, the promoter may contain one or more

30 nucleotide substitutions in the naturally-occurring sequence, as long as such substitutions do not affect specificity of recognition by the DNA-dependent RNA polymerase. The promoter is operably linked to either the DNA sequence encoding the minigenome or to the DNA sequence encoding the miniantigenome to direct the

synthesis of either minigenome RNA or miniantigenome RNA, respectively. Preferably, the promoter directs the synthesis of minigenome RNA.

The minigenome encoded by the cDNA comprises the cis-acting 3'-leader and 5'-trailer sequences from the viral genome that are necessary for its replication and transcription by the viral-encoded RNA-dependent RNA polymerase. For the detection of RNA viruses having segmented genomes, it is believed that cis-acting sequences from any of the segments will make a functional minigenome. These cis-acting sequences flank a negative-sense copy of a reporter gene ORF, which is operably linked to gene-start (GS) and gene-end/polyadenylation (GE) transcription signals that are recognized by the RNA-dependent RNA polymerase. The leader, trailer, GS and GE sequences for any particular negative-strand RNA virus can be found in the scientific literature or can be identified by the skilled artisan using well-known and routine techniques. See, e.g., Weng et al., *Genome Res.* 5:202-207, 1995; Garcia-Sastre et al., *Annu. Rev. Microbiol.* 47:765-790, 1993 and references cited therein. In addition, it is intended that the minigenome can contain modifications in the naturally-occurring leader, trailer, GS and GE sequences that do not destroy replication or transcription of the minigenome or production of reporter gene mRNA in the presence of the negative-strand virus.

Any reporter gene that encodes a detectable product is suitable for use in the present invention. The reporter gene product is preferably one that can easily be assayed for or detected in a cell. One enzyme that has proved to be particularly useful as a reporter gene product is β -galactosidase. Preferably, a bacterial β -galactosidase is used, and most preferably the β -galactosidase from *E. coli* that is encoded by the LacZ gene. β -galactosidase is preferred because of its well-characterized nature and the existence of a variety of methods to detect its presence. Other reporter gene products useful in this invention generally include hydrolases or oxidoreductases and, in particular, such enzymes as β -glucosidase, β -glucuronidase, β -hexosaminidase, luciferase, phospholipase, phosphatase, etc. Green fluorescent protein (GFP) is another reporter gene product useful in the invention.

An ORF encoding β -galactosidase or luciferase is particularly preferred for use in this invention because of the numerous methods known to detect expression of these enzymes and the relative sensitivity of such methods. Among these methods include histochemical assays involving a chromogenic or fluorogenic substrate which

permits detection of β -galactosidase activity by a change in the color of the cell that can be detected macroscopically or microscopically. The use of luciferase provides an enzymatic assay that is more sensitive than the colorimetric or fluorometric β -galactosidase assay. Expression of luciferase may be detected by known
5 luminometric methods using luciferin as the enzyme substrate.

The minigenome or miniantigenome synthesized by the DNA-dependent RNA polymerase should have replication competent termini. These can be generated by appropriate construction of the minigenome-encoding cDNA using methods known in the art. For example, the cDNA can include at least one transcription terminator
10 recognized by the DNA-dependent RNA polymerase in operable linkage with the minigenome or the miniantigenome. More preferably, the cDNA also comprises a self-cleaving ribozyme located between the transcription terminator and the 3'-terminus of the minigenome or miniantigenome. The position of the ribozyme is such that a transcript of the cDNA synthesized by the DNA-dependent RNA polymerase is
15 cleaved by the ribozyme to produce a minigenome RNA or miniantigenome RNA having a replication competent 3'-terminus. Similarly, the cDNA can also contain a self-cleaving ribozyme positioned between the promoter for the DNA-dependent RNA polymerase and the trailer or leader sequence of the minigenome or miniantigenome, respectively, such that the 5'-terminus is replication competent. A
20 number of self-cleaving ribozymes suitable for use in the invention are known in the art. See, e.g., Grosfeld et al., *supra*; Long et al., *FASEB J.* 7:25-30, 1993; Perrotta et al., *Nature* 350:434-436, 1991.

The genetically engineered cell also comprises one or more nucleotide sequences encoding the nucleocapsid proteins that are necessary and sufficient for
25 replication of the minigenome RNA or miniantigenome RNA transcribed from the cDNA. As used herein, "necessary and sufficient for replication" is intended to mean that minigenome RNA or miniantigenome RNA synthesized from the cDNA in the presence of these nucleocapsid proteins is replicated in the absence of the negative-strand virus to be detected while detectable expression of the reporter gene product
30 does not occur until the cell is infected with the negative-strand virus. As used herein, the terms "detectable expression of the reporter gene product" and "dependent upon the presence of the negative-strand virus" mean that the reporter gene product is practically detectable above background levels in the particular diagnostic assay being

used, with background levels referring to measurements taken when the assay is performed in the absence of infectious virus or with genetically-engineered cells that lack the reporter gene.

As demonstrated below, the nucleocapsid proteins necessary and sufficient for replication of an RSV minigenome are the N, P and L proteins. Functional counterparts of these proteins in other negative-strand viruses can be identified from the scientific literature and by using methods known in the art. The combination of nucleocapsid proteins that are necessary and sufficient to replicate the minigenome of any particular negative-strand virus can be determined as described below in Example 3. In brief, T7-expression plasmids are constructed that express each of the nucleocapsid proteins and various combinations of these plasmids are introduced into a vertebrate cell expressing T7 RNA polymerase and containing a minigenome-encoding cDNA. The resulting genetically-engineered cell is incubated for a time sufficient for minigenome replication to proceed and then the cell is mock-infected or infected with the negative-strand virus and expression of the reporter gene product is measured. A combination of nucleocapsid proteins suitable for use in the present invention is one whose pre-infection expression results in cells that lack detectable reporter gene product in the absence of virus but that show efficient expression of the reporter gene product in the presence of the virus. It is intended that any or all of these nucleocapsid proteins may contain amino acid substitutions or deletions in the naturally-occurring sequence as long as the modified protein(s) functions in an equivalent manner as the native protein.

The nucleotide sequences encoding the desired nucleocapsid proteins can be provided by separate polynucleotides cotransfected into the cell or can be provided by a single polynucleotide in which separate transcription units encode the desired nucleocapsid proteins. Expression of mRNAs for the nucleocapsid proteins can be directed by promoters and transcription termination signals recognized by the heterologous DNA-dependent RNA polymerase expressed in the host cell cytoplasm or by polymerase II in the host cell nucleus. In embodiments in which a noncytopathic replicon expresses the DNA-dependent RNA polymerase, the replicon can also contain the nucleotide sequences encoding the nucleocapsid proteins arranged such that the replicon expresses mRNAs for each of the nucleocapsid proteins from a different subgenomic promoter. In a preferred embodiment, the

heterologous DNA-dependent RNA polymerase is T7, T3, or SP6 RNA polymerase and the nucleotide sequences for the desired nucleocapsid proteins are operably linked to the corresponding T7, T3, or SP6 promoter.

In accordance with the method provided by the invention, the above-described
5 genetically-engineered cells can be used to test a sample for the presence of a negative-strand virus. The sample can be any material which can be placed into a fluid or fluid environment and includes biological fluids such as blood, semen, nasopharyngeal swabs, cerebrospinal fluids and the like.

To carry out the method, the genetically engineered cells are first cultured for
10 a period of time sufficient for synthesis of the minigenome or miniantigenome RNA and for expression of the nucleocapsid proteins. It is believed this culture step provides the minigenome RNA in an encapsidated form that is accessible for efficient expression of the reporter gene product by the replicase/transcriptase machinery of the incoming virus. Typically, this first culture period will be between 16 and 24 hours,
15 but shorter or longer culture times may also produce functional, encapsidated minigenome RNA and the culture time suitable for a particular assay can be determined empirically.

The pre-cultured cells are then incubated with the sample in a suitable culture medium for a period of time sufficient for the infectious cycle of the target RNA virus
20 to proceed. If the target virus is in the specimen, it will produce the factor(s) necessary for detectable expression of the reporter gene product. Typically, this incubation period is between 24 and 48 hours, although the optimal length for any particular negative-strand RNA virus should be determined empirically. Expression of the reporter gene product can be detected in the cells, in the culture medium, or in
25 cell extracts.

The invention also provides a kit for detecting a negative-strand RNA virus. The kit is prepared by placing in a container a supply of the above-described genetically-engineered cells sufficient to conduct an assay or a number of assays in accordance with the invention. Preferably, the kit is also provided with reagents
30 necessary for detecting the reporter gene product, placed in separate containers. An instruction manual can also be included in the kit.

The method and cells of this invention are useful for the detection of negative-strand RNA viruses belonging to the following families: *Rhabdoviridae*, *Filoviridae*, *Paramyxoviridae*, *Orthomyxoviridae*, *Bunyaviridae*, and *Arenaviridae*.

The following examples of the present invention are offered by way of illustration
5 and are not to be considered in a limiting sense.

Example 1

This example illustrates that T7 RNA polymerase expressed by a Sindbis virus replicon in baby hamster kidney (BHK) cells is functional.

10 As discussed above, studies of replication and transcription in negative-strand viruses have primarily used a vaccinia virus/T7 RNA polymerase system to supply intracellular T7 RNA polymerase for transcription of a desired gene under the control of the T7 promoter. However, an RNA detection method requiring coinfection with vaccinia virus would not be practicable for clinical diagnostic purposes. Thus, the inventors herein wished to develop a
15 system that would permit the use of stably transformed cells, which would be easier to handle and allow more widespread use.

Recently, noncytopathic Sindbis virus replicons have been developed which contain the gene for puromycin resistance (*pac*) under control of the Sindbis subgenomic RNA promoter (SR19/*pac*) (Frolov et al., *Proc. Natl. Acad. Sci. USA* 93:11371-11377, 1996).
20 Transfection of BHK cells with RNA transcribed *in vitro* from a cDNA of an SR19/*pac* replicon (Fig. 1A) produces a population of puromycin-resistant cells due to constitutive replication and transcription of the noncytopathic replicon and consequent expression of puromycin acetyl transferase. Modification of SR19/*pac* replicons to express an additional foreign gene operably linked to a second subgenomic RNA promoter has been achieved
25 (Lindenbach et al., *J. Virol.* 71:9608-9617, 1997). For example, cells that carry a replicon expressing T7 RNA polymerase (SR19/T7Pol/*pac*) can transcribe a gene under the control of the T7 promoter following transfection of that gene (Agapov, E., et al., *Proc. Natl. Acad. Sci. USA* 95:12989-12994, 1998).

To determine if the SR19/T7pol/*pac* system could be used to express functional T7
30 RNA polymerase in BHK cells, replicon RNAs were transcribed *in vitro* from SR19/*pac* and SR19/T7Pol/*pac* cDNA plasmids (Figs. 1A, 1B) (gift of Eugene Agapov and Charles Rice, Washington University School of Medicine, St. Louis, MO) using SP6 RNA polymerase. The replicon RNAs were electroporated into BHK-21 cells (ATCC) and replicon-containing cell populations were selected for puromycin resistance as described by Lindenbach et al.,
35 *supra*. The SR19/*pac*- and SR19/T7Pol/*pac*-transformed cells were then transfected with the

T7 expression vector pTM1 (a generous gift of Bernard Moss, NIAID, Bethesda, MD) or with pTM1-LacZ, which was generated by cloning the *E. coli lacZ* gene into the *NcoI-BamHI* window of the pTM-1 plasmid, thus placing the *lacZ* gene under control of the T7 promoter. The four groups of recombinant cells were maintained in puromycin-containing medium for 24 hours and then β -galactosidase activity in lysates prepared from these cells was determined as follows.

β -galactosidase activity was measured using the substrate chlorophenolred- β -D-galactopyranoside (CPRG, Boehringer Mannheim, Indianapolis, IN) (final concentration 5 mM) in a 0.2 M potassium phosphate buffer, pH 7.8, with 1 mM $MgCl_2$. Lysates were made in this buffer containing 1% Triton X-100 and 1 mM dithiothreitol. Fifty μ liters of lysate were mixed with 50 μ l of substrate in a microtiter plate well. After incubation for 30 to 120 min at room temperature, the optical density at a wavelength of 562 nm (OD_{562}) was measured with a THERMOmax microplate reader using SOFTmax software (Molecular Devices, Sunnyvale, CA). The assay was shown to be linear up to an OD_{562} of 3.0.

The amount of β -galactosidase activity in the four different groups of cells is shown in Fig. 2 (amounts shown are the mean of duplicate samples). High levels of β -galactosidase activity were detected in cells containing both the SR19/T7Pol/pac replicon and the pTM1-LacZ plasmid, but not in control cells that lacked the T7 RNA polymerase or the *lacZ* gene (Figure 2). Thus, functional T7 RNA polymerase is expressed in BHK cells transfected with SR19/T7pol/pac RNA.

Example 2

This example illustrates that SR19/T7Pol/pac-transformed BHK cells are susceptible to infection by the negative-strand virus RSV and that such RSV-infected cells support replication and transcription of an RSV minigenome.

SR19/T7Pol/pac BHK cells were mock-infected or infected with 10 plaque-forming units (pfu)/cell of RSV (strain A2), which was propagated and titered on HEp-2 cells as described previously (Grosfeld et al., *supra*). Forty-five minutes later, the cells were transfected over a 3-h period with 0.5-1.5 μ g of *in vitro*-synthesized RSV-CAT minigenome RNA, which contains the bacterial *cat* gene in place of all of the viral protein open reading frames (Collins et al, *Proc. Natl. Acad. Sci. USA* 88:9663-9667, 1991). The minigenome RNA was a run-off transcript of 935 nt produced by T7 RNA polymerase of *HgaI*-linearized RSV-CAT cDNA (Collins et al., 1991). Only RSV-infected cells exhibited reporter gene

expression (data not shown), indicating that SR19/T7Pol/pac BHK cells are permissive for RSV infection as well as RSV minigenome replication and expression.

The inventors next asked whether minigenome RNA transcribed *in vivo* in SR19/T7Pol/pac BHK cells was functional. SR19/T7Pol/pac BHK cells were mock-infected or infected with RSV (strain A2) at 10 plaque-forming units (pfu) per cell and 45 min. later were transfected with pC2CAT, a derivative of RSV-CAT cDNA which, *inter alia*, contains a hammerhead-type ribozyme sequence and T7 transcription terminator at the 3' end of the encoded transcript (Grosfeld et al., 1995, *supra*). Ribozyme self-cleavage of the T7-synthesized transcript of pC2CAT leaves a single non-RSV-specific 3'-phosphorylated U residue on the 3' terminus (Grosfeld et al., 1995, *supra*). Under these conditions, RSV-infected cells exhibited a marked increase in CAT activity as compared to CAT activity in mock-infected cells (data not shown).

The results of these experiments indicate that RSV-infected SR19/T7Pol/pac cells can produce a functional RSV minigenome from minigenome RNA or minigenome cDNA that is subsequently transfected into the cell.

Example 3

This example illustrates that preformed minigenome RNA requires RSV nucleocapsid proteins for activity after RSV infection.

The assay initially envisioned by the inventors for detecting RSV in cell culture involved constitutive expression of an RSV minigenome RNA that would be replicated and transcribed after subsequent infection with RSV. To test the feasibility of this protocol, the following experiment was performed.

SR19/T7Pol/pac cells were transfected with pMP210, a cDNA of an RSV minigenome containing a negative-sense copy of the gene for green fluorescent protein (GFP), and 6-24 h later the cells were infected with 10 pfu of RSV. pMP210 was made from pC2Luc cDNA, which contains the luciferase gene substituted for the CAT gene in the above-described pC2CAT construct (Collins et al., *Proc. Natl. Acad. Sci. USA* 93:81-85, 1996). A PCR product containing the Green Lantern Protein gene (Life Technologies, Gaithersburg, MD) flanked by gene start (GS) and gene end (GE) signals was inserted into the BstXI site in the leader of pC2Luc, and the luciferase gene was deleted. In addition, a 5' hammerhead ribozyme preceded by a unique *AvrII* site (Altschuler et al., *Gene* 122:85-90, 1992) was inserted just before the trailer sequence. The pC2 3' hammerhead ribozyme was replaced with a PCR product containing the antigenomic hepatitis delta virus ribozyme (Perrotta et al., *Nature* 350:434-436, 1991) followed by a unique *XhoI* site.

Under these conditions, no RSV-dependent GFP expression was observed. These negative results suggest that minigenome RNA has a very short half-life in uninfected BHK cells or is in some manner not accessible to or recognized by the incoming RSV replicase and/or transcriptase. Thus, a viable assay for detecting RSV infection using an RSV minigenome requires a strategy for stabilizing the minigenome RNA and/or making it replication competent.

It has been shown that RSV N, P, and L proteins are necessary and sufficient for replication of RSV minigenomes and that expression of the N, P, and L proteins leads to transcription of RSV mRNA, although not necessarily full-length mRNA (Grosfeld et al., *supra*; Yu et al., *supra*. In addition, expression of only the N and P proteins leads to encapsidation of RNA minigenomes and this is increased 10 to 50-fold by the addition of the L protein (Atreya et al., *J. Virol.* **72**:1452-1461, 1998). Based on these results, the inventors investigated whether a minigenome expressed in the presence of the N, P, and L proteins would be accessible to transcription of reporter gene mRNA when the cells were subsequently infected with RSV.

SR19/T7Pol/pac cells or SR19/pac cells were transfected with 1 μ g of pMP210LacZ, a lacZ-containing minigenome cDNA, with or without 0.2 μ g each of the T7 expression plasmids pTM1-N, pTM1-P, and pTM1-L described in Grosfeld et al., *supra*. pMP210LacZ was made by replacing the ORF of GFP in pMP210 with the ORF of lacZ using the unique XbaI and XmaI sites. Twenty hours after transfection the cells were trypsinized and plated in 24 well dishes and six hours later infected with RSV (5 pfu/cell) or mock-infected. At 36 hrs after infection, the cells were lysed and assayed for β -galactosidase as described above. The results, which are the mean of duplicate samples, are shown in Figure 3A.

Significant induction of β -galactosidase activity by RSV infection was observed in cells expressing both T7 RNA polymerase and the RSV nucleocapsid proteins N, L and P. These cells are illustrated in Figure 7. Notably, β -galactosidase activity in the corresponding mock-infected cells was not significantly higher than β -galactosidase activity in mock-infected cells lacking T7 (background) or in mock-infected cells lacking N, L and P.

To assess the relative requirement for each of the nucleocapsid proteins, various combinations of the N, L and P expression plasmids (0.2 μ g each) were cotransfected with the minigenome cDNA (pMP210LacZ at 1 μ g) into SR19/T7Pol/pac cells and the assay was carried out as above. The results, which are the mean of triplicate samples \pm the standard deviation, are shown in Fig. 3B. RSV-induced β -galactosidase activity was significant only in the presence of all three nucleocapsid proteins. In addition, the amount of activity was unchanged when the assay was carried out after 10 passages of these minigenome-expressing cells.

When evaluated by histochemical staining as described in U.S. Patent 5,418,132, β -galactosidase-positive infected cells were observed only in the presence of the nucleocapsid proteins: a few stained infected cells per well were observed with N alone or with N plus P, while many stained infected cells were consistently observed when N, P, and L were all expressed (data not shown). Significantly, no stained cells were observed in the absence of RSV with or without the nucleocapsid proteins. The requirement for all three nucleocapsid proteins in this assay suggests that encapsidation and replication of the minigenome RNA are necessary for efficient reporter gene expression by the incoming RSV. In addition, the lack of β -galactosidase activity in mock-infected cells expressing all three nucleocapsid proteins indicates that detectable expression of β -galactosidase is dependent upon some factor(s) provided by RSV infection, probably the M2-1 protein.

Example 4

This example illustrates that the level of expression of the minigenome reporter gene is dependent on the input concentration of RSV.

Approximately 2×10^6 SR19/T7Pol/pac cells in six 9.5 cm² wells were transfected with pMP210LacZ (1 μ g) and pTM1-N, P, and L (0.2 μ g each). After 18 h the transfected cells were trypsinized, pooled and plated into all the wells of a 48-well plate. Six h later approximately 10^5 cells were either (A) infected with increasing volumes of a two-fold dilution of a stock RSV (titer 2×10^7 pfu/ml assayed on HEp2 cells) and β -galactosidase activity assayed 36 hr later or (B) infected with either 0.5 microliters (low multiplicity of infection; LMOI) or 25 microliters (high multiplicity of infection; HMOI) of the same stock virus and β -galactosidase activity assayed at the indicated times after infection. The results of the dose response assay are shown in Fig. 4A (expressed as the mean of triplicate samples \pm the standard deviation) and the results of the time course assay are shown in Fig. 4B (the mean of duplicate samples).

The assay exhibited a relatively linear dose-response for RSV over a range of greater than one log₁₀ (Figure 4A). The RSV-induced expression exhibited a plateau effect above an inoculum equivalent to a multiplicity of infection of approximately 0.6 pfu/cell (based on HEp2 cells). These results indicate that this assay could be used in place of a plaque assay to quantitate RSV stocks.

When β -galactosidase in these minigenome-expressing cells was measured as a function of time after infection, β -galactosidase activity following a LMOI infection (0.2 pfu/cell) reached the same level as that following a HMOI infection (10 pfu/cell), but with a

delay of approximately 48 h (Fig. 4B). This suggests that RSV replicates and spreads within these cell cultures and that the duration of an infectious cycle is approximately 48 h.

Example 5

5 This example illustrates that RSV-induced expression of the minigenome reporter gene is sensitive to ribavirin.

Ribavirin inhibits *in vitro* replication of many RNA viruses including RSV (Sidwell et al., *Pharmacol Ther* 6:123-146, 1979). Depending on the assay used, the 50% effective dose (ED₅₀) of ribavirin for RSV has been reported to be around 5 µg/ml (Chiba et al., *Biol*
10 *Pharm Bull* 18:1081-1083, 1995). To determine if ribavirin affects the detection of RSV by a minigenome assay, SR19/T7Pol/pac cells were cotransfected with pMP210LacZ, pTM1-N, pTM1-P, and pTM1-L as described in Example 3 and then infected with RSV (5 pfu/cell) in the presence of 1, 5, 50 or 200 µg/ml of ribavirin. As shown in Fig. 5A, β-galactosidase activity was reduced to near background levels by as little as 1 µg/ml ribavirin (Fig. 5A).
15 When this dose response experiment was repeated at subinhibitory concentrations between 0 and 400 ng/ml ribavirin, the ED₅₀ of ribavirin was determined to be less than 100 ng/ml (Fig. 5B).

Treatment of the cells with ribavirin (1 µg/ml) from the time of transfection of the RSV cDNA plasmids until the time of RSV infection had no effect on RSV-dependent β-
20 galactosidase expression (data not shown). This suggests that under the conditions in which this assay was performed, ribavirin primarily affects RSV replication and transcription rather than the Sindbis replicon or T7 polymerase transcription and than any minigenome replication mediated by the RSV L protein occurring prior to replication of incoming RSV must be relatively insensitive to ribavirin.

25 Example 6

This example illustrates that RSV-induced β-galactosidase activity in minigenome-expressing cells is sensitive to neutralizing antibody to RSV.

To demonstrate that the RSV-induced β-galactosidase activity in this system requires infectious virus, and can, therefore, be blocked by neutralizing antibody, we incubated RSV
30 with different dilutions of a sample of pooled human gamma globulin. RSV is a highly prevalent human pathogen and most adults have been infected at least once (Collins et al., Respiratory Syncytial Virus, pp. 1313-1350 in *Fields Virology*, B. N. Fields, D. M. Knipe and P. M. Howley, eds., Lippencott-Raven, Philadelphia, 1996). Therefore, pooled human gamma globulin has significant neutralizing activity against RSV. The globulin-treated virus
35 was used to infect SR19/T7Pol/pac cells that had previously been transfected with

pMP210LacZ and pTM1-N, pTM1-P, and pTM1-L. As shown in Figure 6, β -galactosidase activity was inversely proportional to the amount of dilution of the gamma globulin sample.

Example 7

5 This example illustrates the specificity of RSV minigenome-expressing cells in detecting RSV in patient samples.

 SR19/T7Pol/pac cells that had previously been transfected with pMP210LacZ and pTM1-N, pTM1-P, and pTM1-L were incubated with two patient specimens, previously determined to contain RSV, or with patient specimens known to contain influenza A,
10 influenza B, and parainfluenza types 1 and 2. Induction of β -galactosidase activity was determined by histochemical staining.

 Incubation with the RSV-containing specimens produced significant β -galactosidase, while specimens containing the other negative-stranded viruses did not induce any positively stained cells (data not shown). Thus, the assay is specific for RSV.

15 The sensitivity of this assay can be enhanced by increasing the time of infection before assaying for β -galactosidase activity. At 36 h after infection, 3000 pfu could be detected; at 5 days after infection even an inoculum of less than 10 pfu gave a reading several-fold above background.

 In view of the above, it will be seen that the several advantages of the
20 invention are achieved and other advantageous results attained.

 As various changes could be made in the above methods and compositions without departing from the scope of the invention, it is intended that all matter contained in the above description and shown in the accompanying drawings shall be interpreted as illustrative and not in a limiting sense.

25 All references cited in this specification, including patents and patent applications, are hereby incorporated by reference. The discussion of references herein is intended merely to summarize the assertions made by their authors and no admission is made that any reference constitutes prior art. Applicants reserve the right to challenge the accuracy and pertinency of the cited references.

What is Claimed is:

1. A genetically engineered cell for detecting a negative-strand RNA virus, the cell comprising:
 - a polynucleotide encoding a DNA-dependent RNA polymerase;
 - a cDNA comprising a minigenome and a miniantigenome of the negative-strand
 - 5 RNA virus operably linked to a promoter for the DNA-dependent RNA polymerase, wherein the miniantigenome comprises a nucleotide sequence encoding a reporter gene product, and wherein expression of the reporter gene product is dependent upon the presence of the negative-strand RNA virus; and
 - one or more nucleotide sequences encoding each of the nucleocapsid proteins of
 - 10 the negative-strand RNA virus which are necessary and sufficient for replication of minigenome RNA or miniantigenome RNA synthesized by the DNA-dependent RNA polymerase.
2. The genetically engineered cell of claim 1, wherein the DNA-dependent RNA polymerase is T7 RNA polymerase, T3 RNA polymerase or SP6 RNA polymerase.
3. The genetically engineered cell of claim 2, wherein the polynucleotide is integrated into the nucleus and comprises a polymerase II promoter operably linked to a nucleotide sequence encoding the DNA-dependent RNA polymerase.
4. The genetically engineered cell of claim 2, wherein the polynucleotide comprises a noncytopathic positive-strand virus replicon.
5. The genetically engineered cell of claim 4, wherein the noncytopathic positive-strand virus replicon comprises a noncytopathic flavivirus replicon, a noncytopathic alphavirus replicon, a noncytopathic nodavirus replicon, a noncytopathic coronavirus replicon, or a noncytopathic astrovirus replicon.
6. The genetically engineered cell of claim 5, wherein the negative-strand RNA virus is an orthomyxovirus, a paramyxovirus, a rhabdovirus, a filovirus, a bornavirus or a bunyavirus.
7. The genetically engineered cell line of claim 6, wherein the negative-strand RNA virus is selected from the group consisting of: human parainfluenza virus type 1, human parainfluenza virus type 2, human parainfluenza virus type 3, human

parainfluenza virus type 4, human influenza virus, human respiratory syncytial virus
5 (RSV), measles virus, mumps virus, rabies virus, ebola virus and hanta virus.

8. The genetically engineered cell line of claim 7, wherein the reporter gene product is β -galactosidase, chloramphenical acetyl transferase, luciferase, alkaline phosphatase, green fluorescent protein or β -glucuronidase.

9. The genetically engineered cell line of claim 8, wherein the negative-strand RNA virus is RSV, the DNA-dependent RNA polymerase is T7 RNA polymerase, the reporter gene product is β -galactosidase, and the nucleocapsid proteins necessary and sufficient for replication of the minigenome are the RSV proteins N, P and L which are encoded by three
5 T7 expression plasmids.

10. A method for detecting a negative-strand RNA virus in a sample, the method comprising:

- (a) providing a genetically engineered cell which comprises
 - a polynucleotide encoding a DNA-dependent RNA polymerase;
 - 5 a cDNA comprising a minigenome and a miniantigenome of the negative-strand RNA virus operably linked to a promoter for the DNA-dependent RNA polymerase, wherein the miniantigenome comprises a nucleotide sequence encoding a reporter gene product, and wherein expression of the reporter gene product is dependent upon the presence of the negative-strand RNA virus; and
 - 10 one or more nucleotide sequences encoding each of the nucleocapsid proteins of the negative-strand RNA virus which are necessary and sufficient for replication of minigenome RNA or miniantigenome RNA synthesized by the DNA-dependent RNA polymerase;
- (b) culturing the cell for a time sufficient to synthesize the minigenome RNA or
15 the miniantigenome RNA and to express the nucleocapsid proteins;
- (c) incubating the cell with the sample; and
- (d) detecting expression of the reporter gene product.

11. The method of claim 10, wherein the DNA-dependent RNA polymerase is T7 RNA polymerase, T3 RNA polymerase or SP6 RNA polymerase.

12. The method of claim 11, wherein the polynucleotide is integrated into the nucleus and comprises a polymerase II promoter capable of causing transcription of the polynucleotide.

13. The method of claim 11, wherein the polynucleotide comprises a transcript of a noncytopathic positive-strand virus replicon.

14. The method of claim 13, wherein the noncytopathic positive-strand virus replicon comprises a noncytopathic flavivirus replicon, a noncytopathic alphavirus replicon, a noncytopathic nodavirus replicon, a noncytopathic coronavirus replicon, or a noncytopathic astrovirus replicon.

15. The method of claim 14, wherein the negative-strand RNA virus is an orthomyxovirus, a paramyxovirus, a rhabdovirus, a filovirus, a bornavirus or a bunyavirus.

16. The method of claim 15, wherein the negative-strand RNA virus is selected from the group consisting of: human parainfluenza virus type 1, human parainfluenza virus type 2, human parainfluenza virus type 3, human parainfluenza virus type 4, human influenza virus, human respiratory syncytial virus (RSV), measles virus, mumps virus, rabies virus, ebola virus and hanta virus.

17. The method of claim 16, wherein the reporter gene product is β -galactosidase, chloramphenicol acetyl transferase, luciferase, alkaline phosphatase, or green fluorescent protein.

18. The method of claim 17, wherein the negative-strand RNA virus is RSV, the DNA-dependent RNA polymerase is T7 RNA polymerase, the reporter gene product is β -galactosidase, and the nucleocapsid proteins necessary and sufficient for replication of the minigenome are the RSV proteins N, P and L which are encoded by three T7 expression plasmids.

19. A kit for detecting a negative-strand RNA virus sample in a sample, the kit comprising:

a supply of genetically engineered cells which comprise:

a polynucleotide encoding a DNA-dependent RNA polymerase;

5 a cDNA comprising a minigenome and a miniantigenome of the negative-strand RNA virus operably linked to a promoter for the DNA-dependent RNA polymerase, wherein the miniantigenome comprises a nucleotide sequence encoding a reporter gene product, and wherein expression of the reporter gene product is dependent upon the presence of the negative-strand RNA virus; and

10 one or more nucleotide sequences encoding each of the nucleocapsid proteins of the negative-strand RNA virus which are necessary and sufficient for replication of minigenome RNA or miniantigenome RNA synthesized by the DNA-dependent RNA polymerase.

20. The kit of claim 19, further comprising a supply of reagents necessary to detect expression of the reporter gene product.

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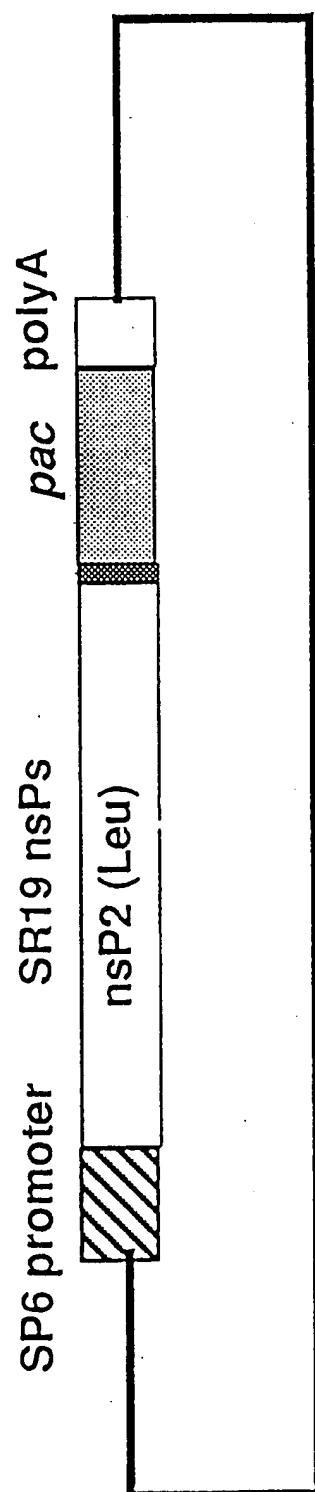


FIGURE 1A

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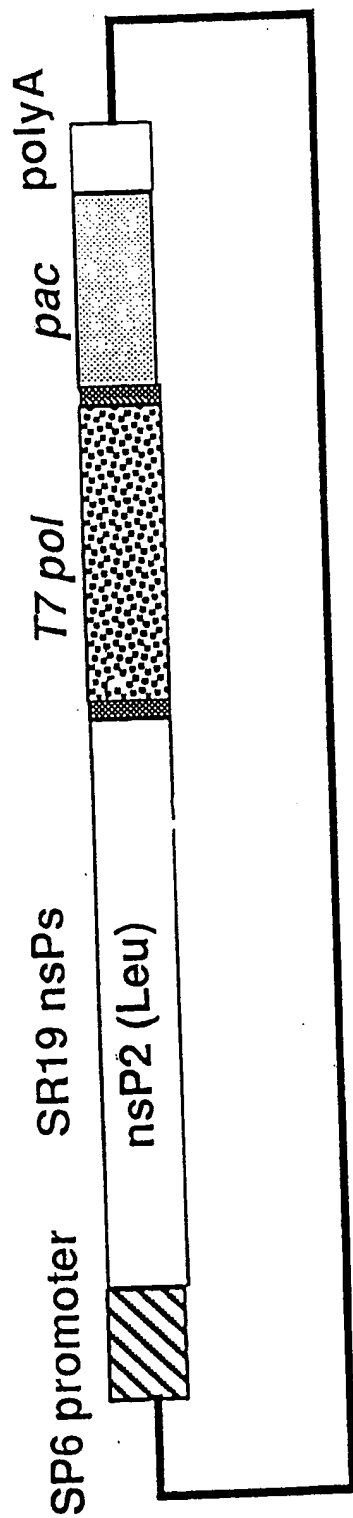


FIGURE 1B

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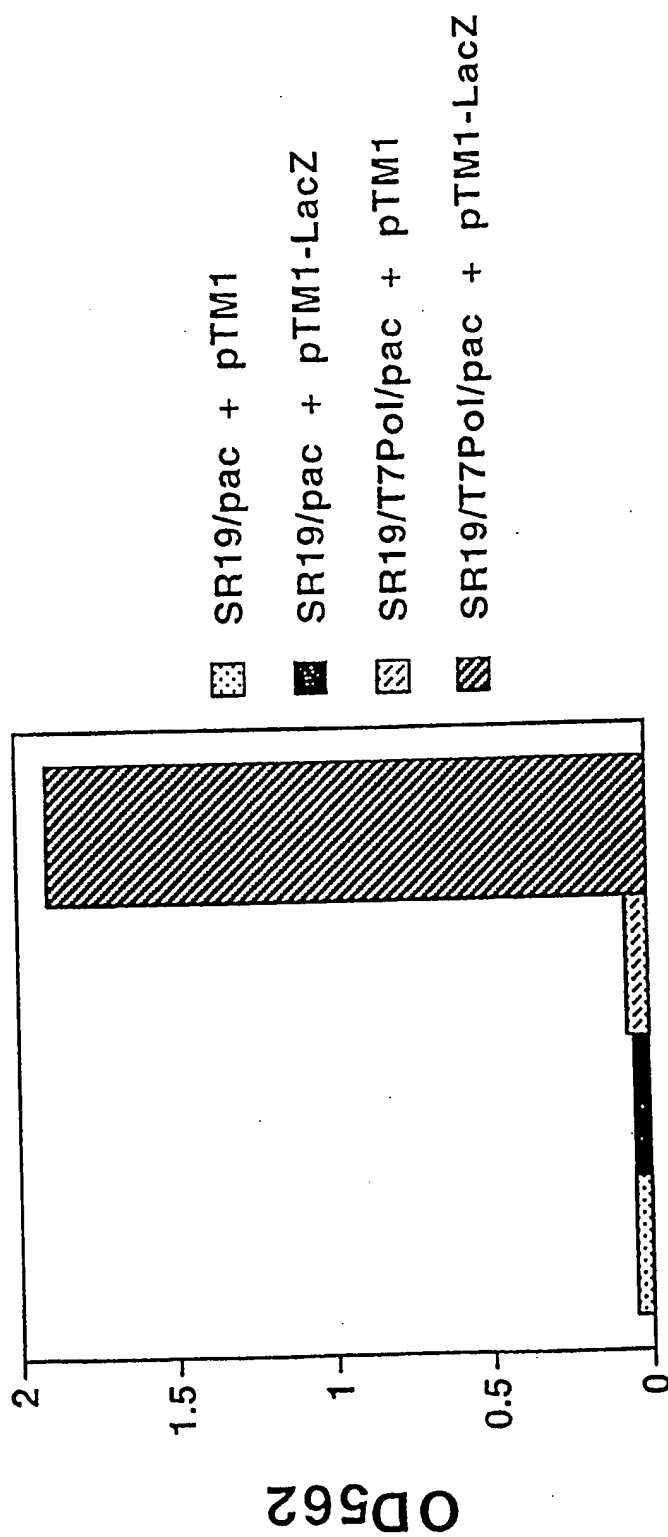


FIGURE 2

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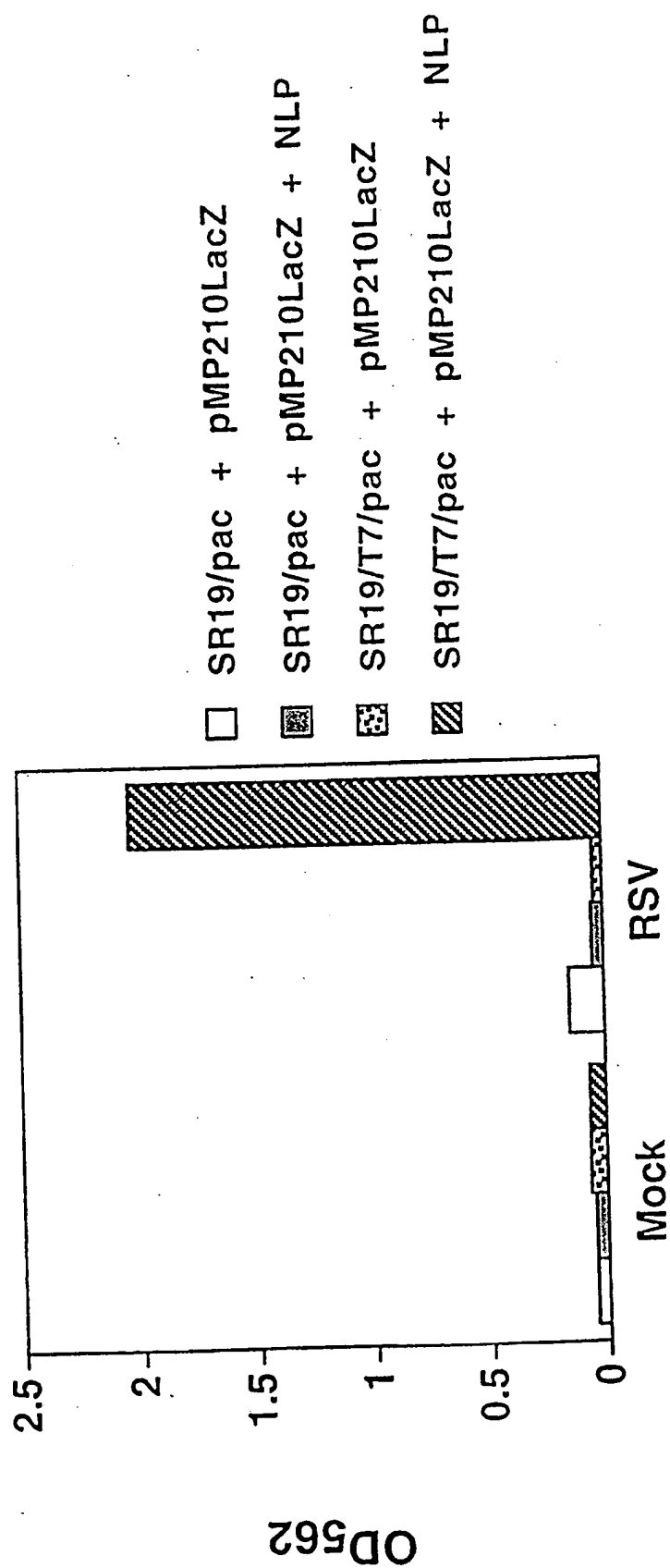


FIGURE 3A

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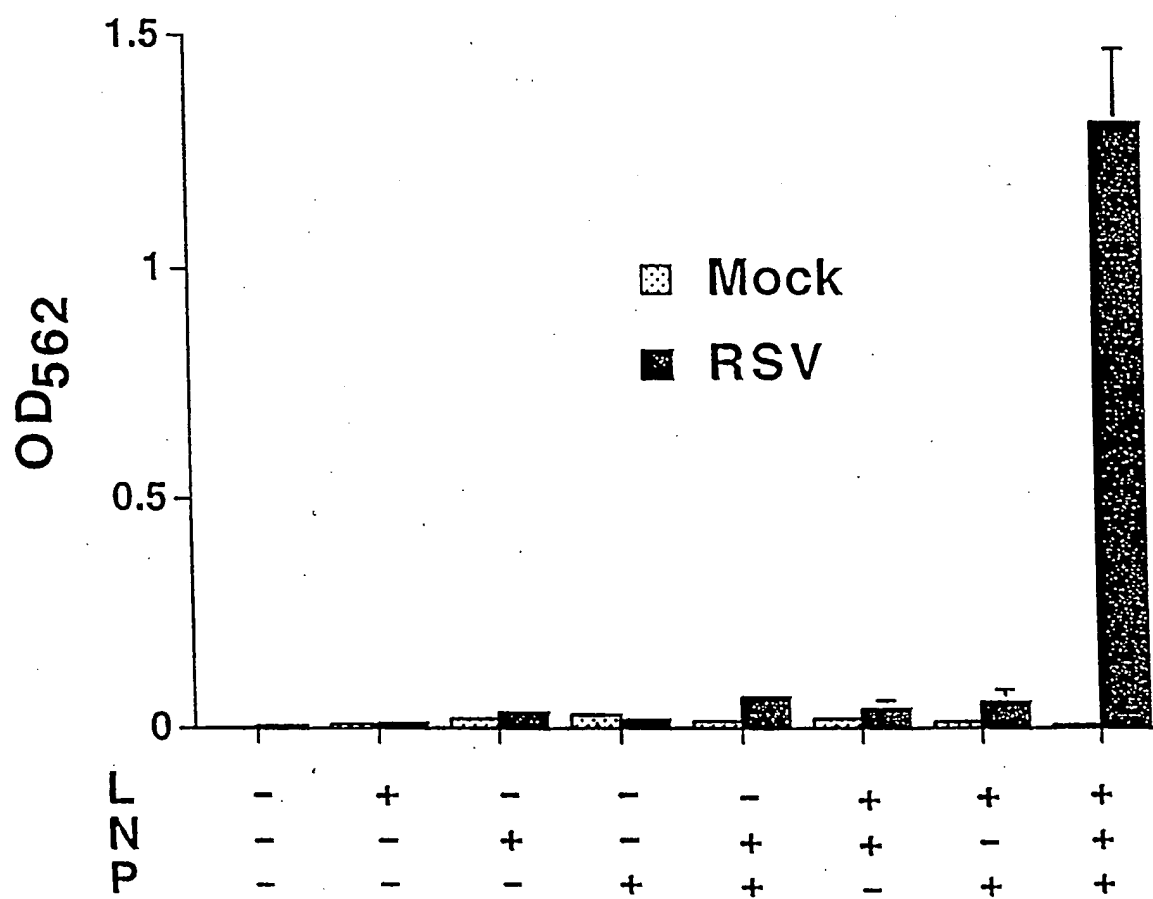


FIGURE 3B

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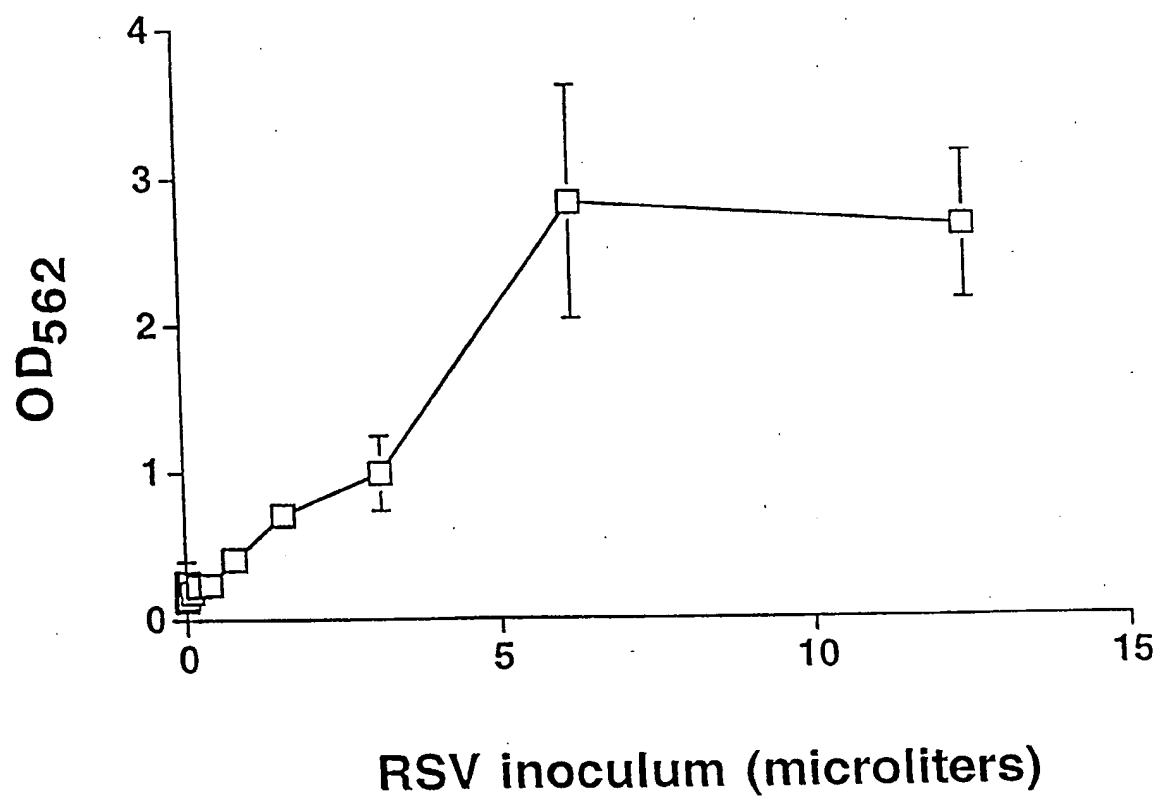


FIGURE 4A

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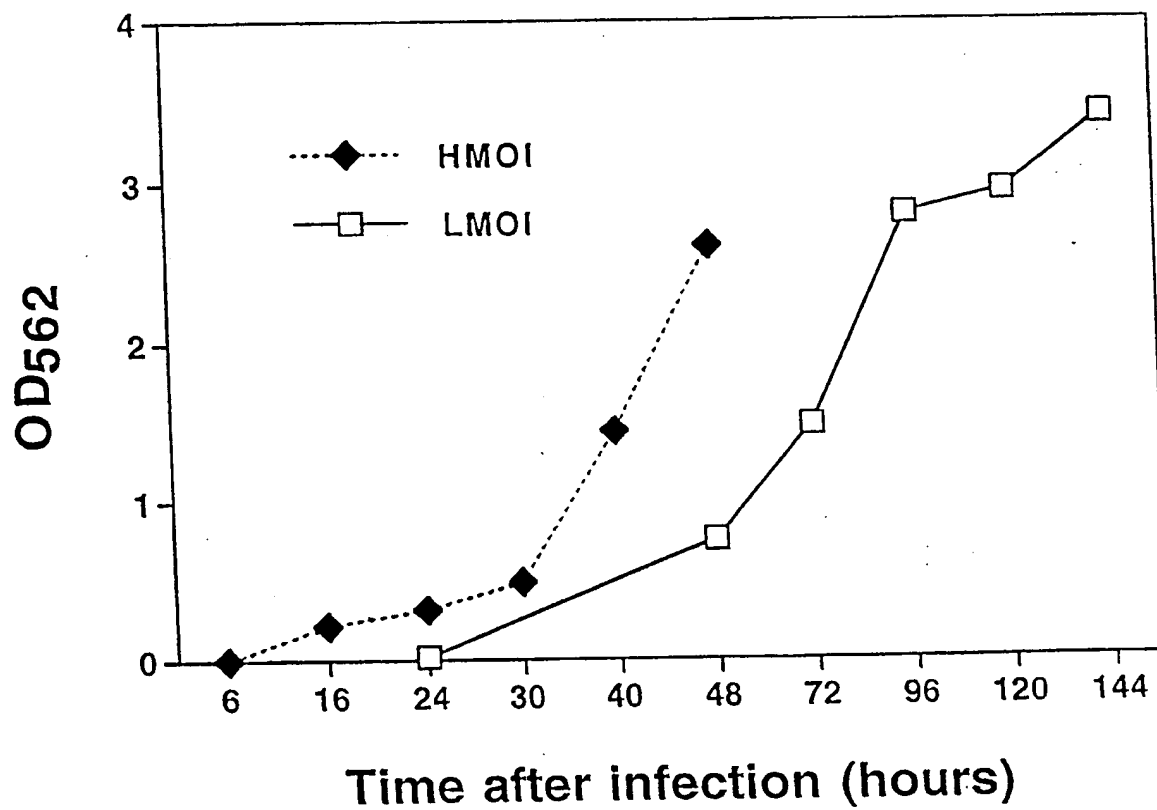


FIGURE 4B

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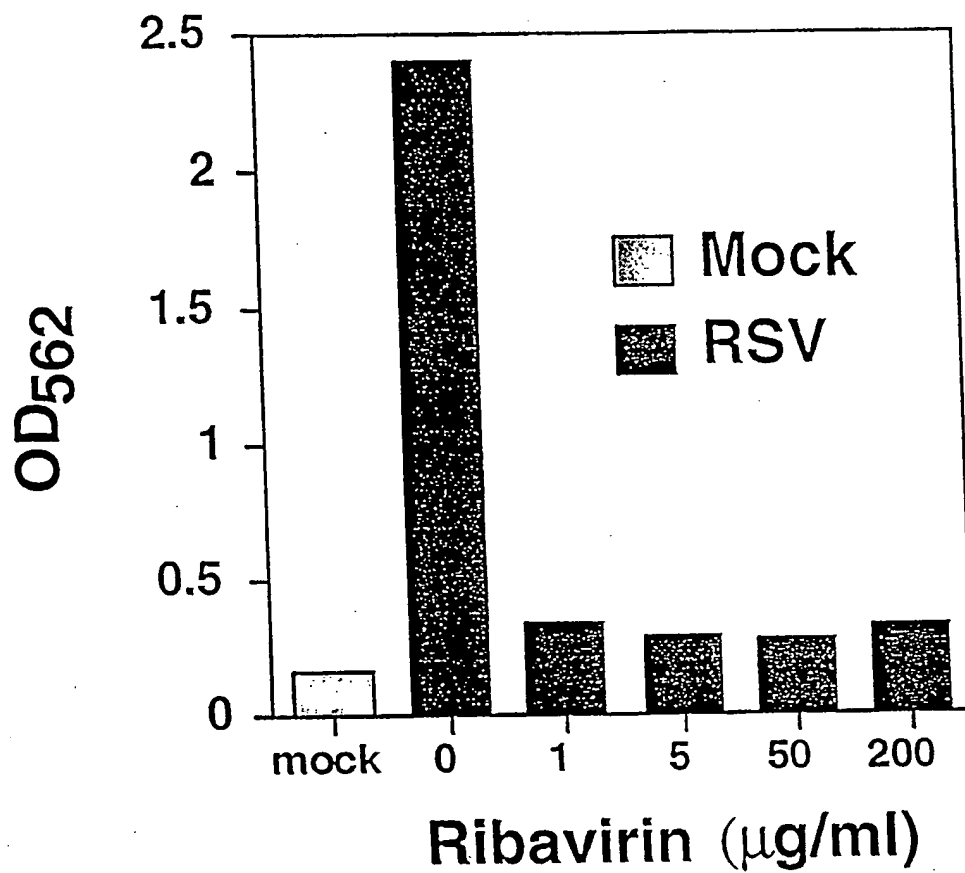


FIGURE 5A

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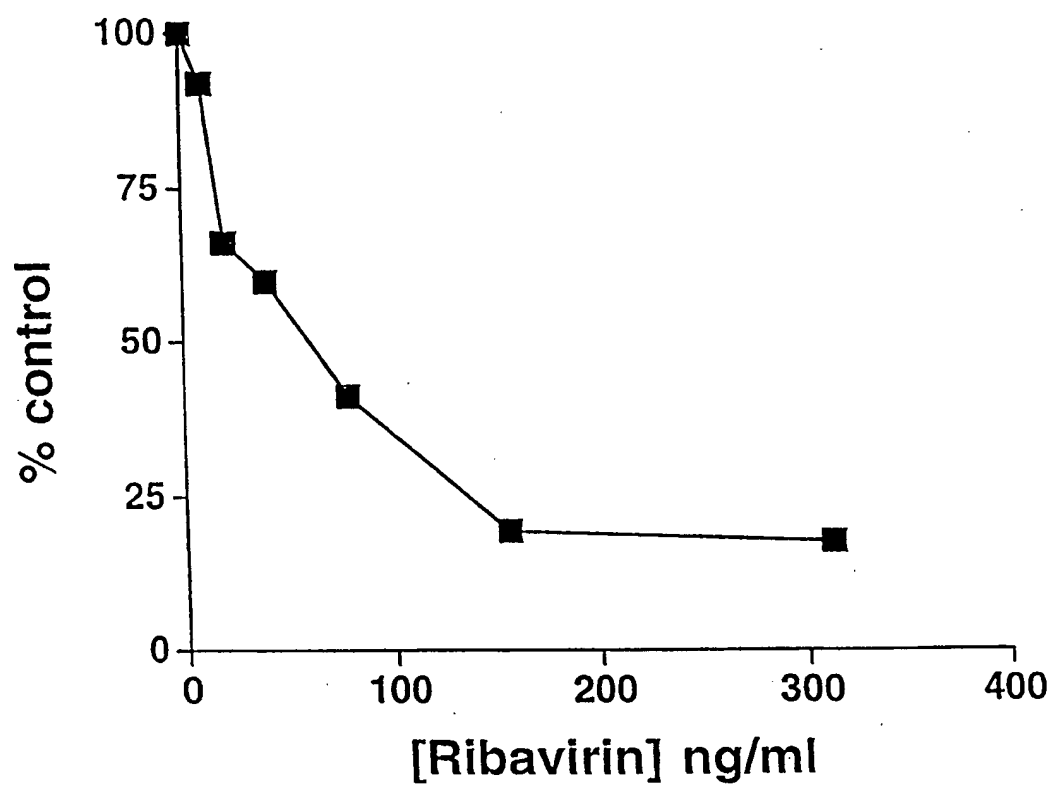


FIGURE 5B

10/11

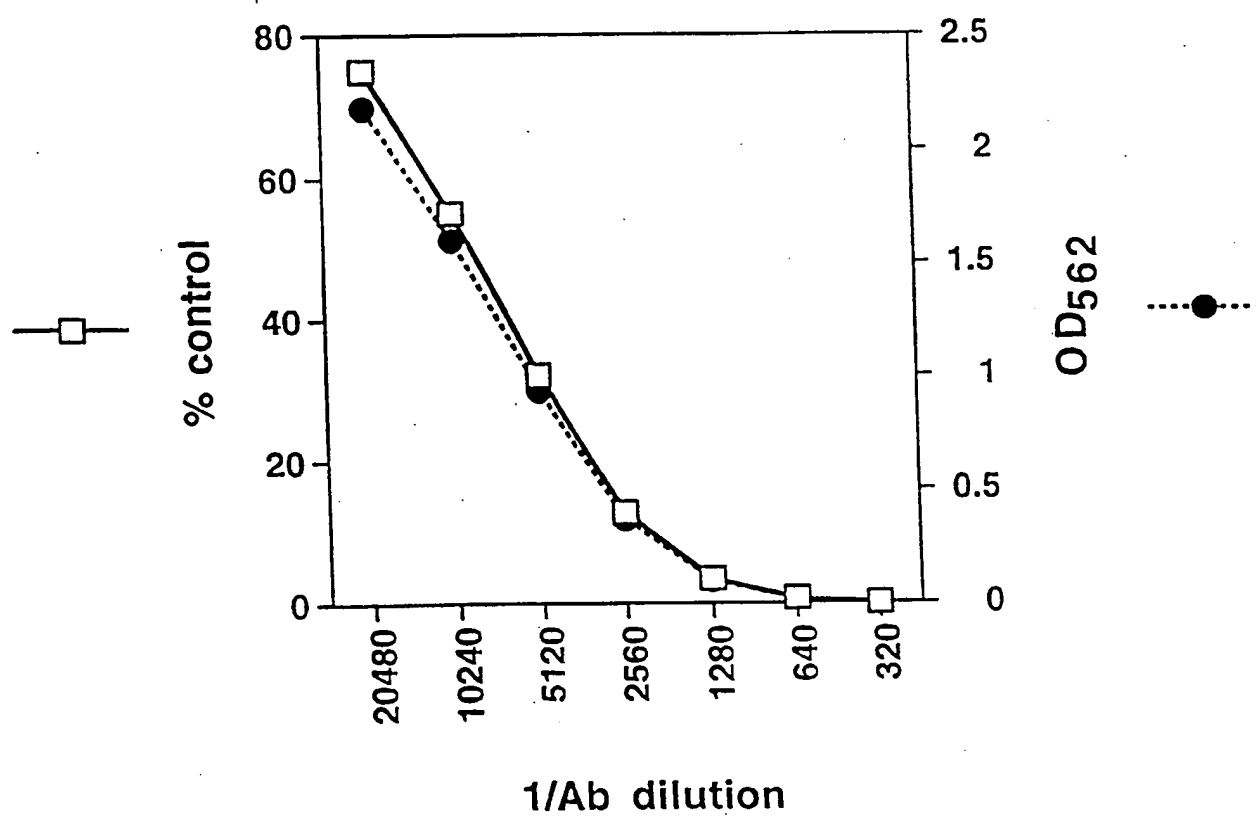


FIGURE 6

11/11

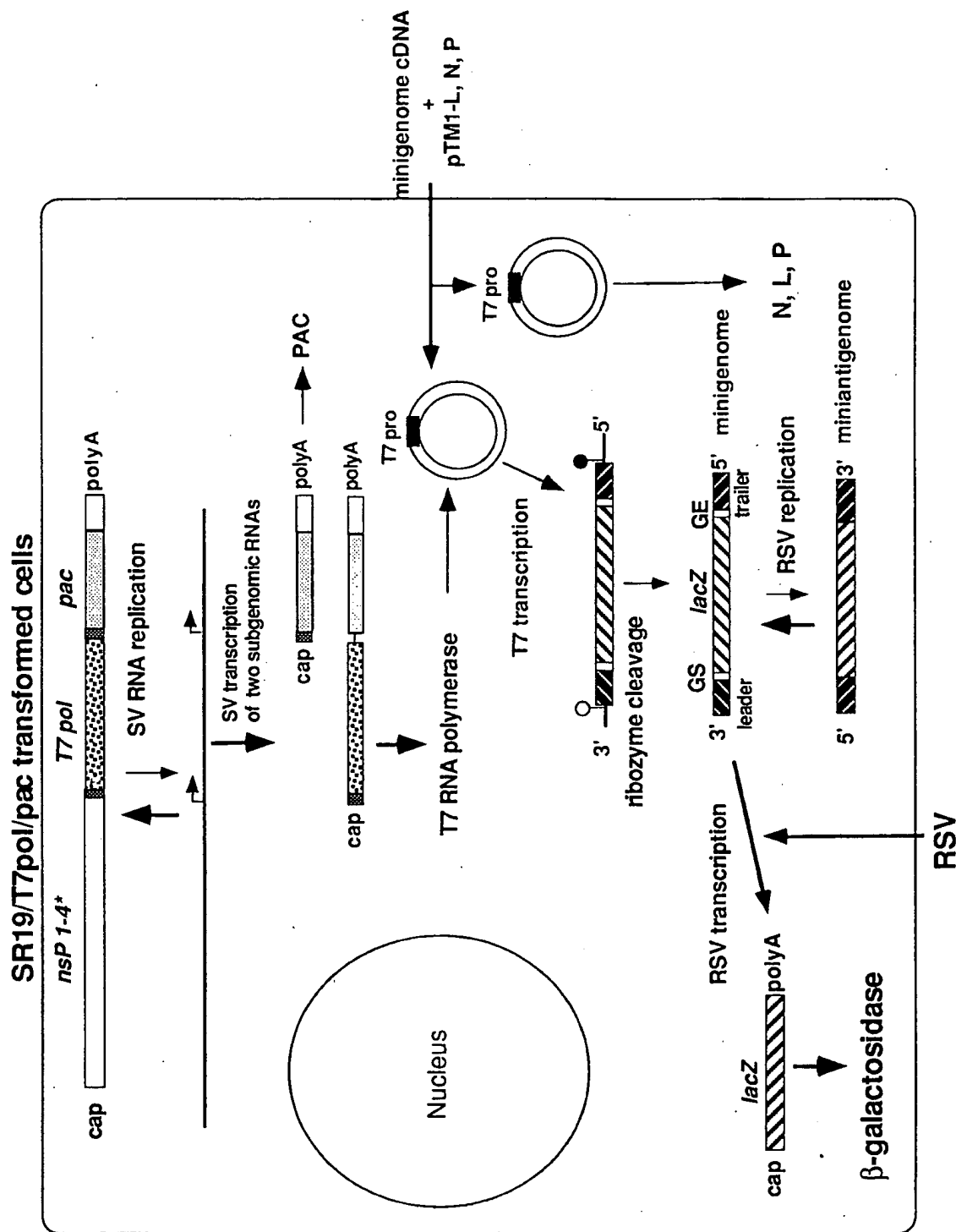


FIGURE 7

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US99/25390

A. CLASSIFICATION OF SUBJECT MATTER

IPC(7) :C12Q 1/70; C12N 5/10

US CL :435/5, 325

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 435/5, 325, 975

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

Please See Extra Sheet.

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X,P	OLIOVO et al. Detection and Quantitation of Human Respiratory Syncytial Virus (RSV) Using Minigenome cDNA and a Sindbis Virus Replicon: A Prototype Assay for Negative-Strand RNA Viruses. Virology. 10 November 1998, Vol. 251, No. 1, pages 198-205, see entire document.	1-20
A	OLIVO, P.D. et al. Transgenic Cell Lines for Detection of Animal Viruses. Clinical Microbiology Reviews. July 1996, Vol. 9, No. 3, pages 321-334, see entire document, especially page 331.	1-20

☐ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

* Special categories of cited documents:	*T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
A document defining the general state of the art which is not considered to be of particular relevance	*X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
E earlier document published on or after the international filing date	*Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
L document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	*G* document member of the same patent family
O document referring to an oral disclosure, use, exhibition or other means	
P document published prior to the international filing date but later than the priority date claimed	

Date of the actual completion of the international search

24 JANUARY 2000

Date of mailing of the international search report

08 FEB 2000

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INTERNATIONAL SEARCH REPORT

International application No.

PCT/US99/25390

B. FIELDS SEARCHED

Electronic data bases consulted (Name of data base and where practicable terms used):

Medline, Derwent Biotechnology, West Derwent WPI, STN USPATFULL, East USPATFULL. Search terms: detect?, transgenic?, negative, nucleocapsid?, orthomyx?, paramyx?, rhabdo?, filo? borna?, bunya?, parainfluenza?, influenza, influenzavir?, rsv, measles, mumps, rabies, ebola, hanta?



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<p>(54) Title: DETECTION OF NEGATIVE-STRAND RNA VIRUSES (57) Abstract A diagnostic assay for detecting a negative-strand RNA virus in a sample and a genetically engineered cell for use in the assay are disclosed. The cell expresses a heterologous DNA-dependent RNA polymerase that synthesizes a minigenome or miniantigenome of the RNA virus from a cDNA template present in the cell. The cell also expresses the nucleocapsid proteins of the negative-strand virus that are necessary for replication of the minigenome or miniantigenome. Infection of the cell by the negative-strand virus results in expression of a reporter gene product encoded by the miniantigenome.</p>		

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DETECTION OF NEGATIVE-STRAND RNA VIRUSES

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5 invention.

Background of the Invention

(1) Field of the Invention

This invention generally relates to the field of virology and, more particularly,
10 to a method for detecting a negative-strand RNA virus in a biological specimen and genetically engineered cells for use in the method.

(2) Description of Related Art

Numerous negative-strand RNA viruses are pathogenic to humans and other animals. Examples of human diseases caused by negative-strand viruses include
15 mumps, measles, pneumonia, bronchitis, influenza, infectious croup, rabies, ebola hemorrhagic fever, marburg hemorrhagic fever, and LaCrosse encephalitis. Thus,

detection of negative-strand viruses in biological specimens is important clinically and for various research purposes.

Although there is considerable diversity in the genomic structure and biological properties of negative-strand RNA viruses, the RNA replication and transcription strategies of these viruses have common features. As with all RNA viruses, negative-strand RNA viruses express an RNA-dependent RNA polymerase and other RNA replicase and transcriptase factors necessary to transcribe their mRNA and replicate their genomes (Olivo, P.D., *Clin. Microb. Rev.* 9:321-334, 1996). In the virion of a negative-strand virus, the RNA polymerase forms a complex with the nucleocapsid protein and the genomic RNA. Once the virion enters the cell and begins to uncoat, the virion RNA polymerase transcribes subgenomic mRNAs from the genomic RNA. The RNA polymerase also synthesizes full-length positive-strand replicative-intermediate RNA (antigenome), which is used as a template for making many copies of negative-stranded genomic RNA that then are used as templates for secondary transcription of additional viral mRNAs.

The study of negative-strand viruses has been complicated by the fact that the RNA-dependent RNA polymerase uses as template only RNA associated with virus-specific nucleocapsid proteins; thus, naked genomic RNA transfected into a cell is not replicated (Bukreyev et al., *J. Virol.* 70:6634-6641, 1996; Olivo, *supra*). Recently, however, a number of laboratories have described advances in the genetic manipulation of various negative-strand RNA viruses. For example, infectious rabies virus, Sendai virus, vesicular stomatitis virus, measles virus, and respiratory syncytial virus (RSV) have been reportedly recovered by using T7 RNA polymerase to generate a full-length antigenomic transcript from a cDNA of the genome in the cell cytoplasm together with the viral proteins necessary for assembly of a nucleocapsid and for RNA replication and transcription (Schnell et al., *EMBO J* 13:4195-4203, 1994; Garcin et al., *EMBO J* 14:6087-6094, 1995; Lawson et al., *Proc. Natl. Acad. Sci. USA* 92:4477-4481, 1995; Radecke et al., *EMBO J* 14:5773-5784, 1995; Whelan et al., *Proc. Natl. Acad. Sci. USA* 92:8388-8392, 1995; Collins et al., *Proc. Natl. Acad. Sci. USA* 92:11563-11567, 1995). In these reports, the T7 RNA polymerase was provided to the cell cytoplasm by infection with a recombinant vaccinia virus containing the gene for T7 RNA polymerase.

Other studies investigating viral protein function in replication and transcription of negative-strand RNA viruses have employed "minigenomes", in which some or all of the viral protein-coding sequences are replaced with a reporter gene flanked by *cis*-acting elements necessary for replication and transcription. See, 5 e.g., Grosfeld et al., *J. of Virol.* 69:5677-5686, 1995; Conzelmann et al., *J. Virol.* 68:713-719, 1994; De et al., *Virol.* 196:344-348, 1993; Dimock et al., *J. Virol.* 67:27722-2778, 1993. In this approach, a cDNA is constructed in which the minigenome is operably linked to a promoter for a bacteriophage RNA polymerase, and the minigenome cDNA is transfected into cells and transcribed by the 10 bacteriophage RNA polymerase in the presence of various viral proteins supplied by cotransfected plasmids, whose expression is also driven by the bacteriophage RNA polymerase, which is supplied by infection with a recombinant vaccinia virus. Induction of reporter gene expression indicates the transfected cell is expressing all the viral proteins needed to replicate and transcribe the minigenome.

15 Using this approach to study replication and transcription in RSV, the major nucleocapsid protein (N), the nucleocapsid-associated phosphoprotein (P), and the polymerase L protein were identified as sufficient to replicate the minigenome (Grosfeld et al., *supra*; Yu et al., *J. Virol.* 69:2412-2419, 1995). Coexpression of the N, P, and L proteins with a minigenome containing the *cat* reporter gene also resulted 20 in synthesis of full-size and incomplete CAT mRNA species and abundant expression of CAT (Grosfeld et al., *supra*). In contrast, efficient synthesis of full-length mRNA was observed when cells containing the minigenome cDNA and the N, P and L expression plasmids were coinfectd with the T7-expressing vaccinia virus and RSV, i.e., the minigenome RNA and the N, P and L proteins were synthesized by T7 RNA 25 polymerase in the presence of infectious RSV (Grosfeld et al., *supra*). A later study demonstrated that expression of an additional RSV protein, the transcription elongation factor M2-1, was required for fully processive sequential transcription in this T7-expression plasmid complementation system (Collins et al., *Proc. Natl. Acad. Sci. USA* 93:81-85, 1996).

30 In U.S. Patent No. 5,591,579, a genetically engineered cell line and method for detecting positive-strand RNA viruses was described. The cell line is stably transformed with a cDNA copy of a structurally defective RNA virus genome which contains (1) the *cis*-acting sequences of the RNA virus genome which are necessary

for replication and transcription by trans-acting enzymes from the RNA virus and (2) a reporter gene. The cDNA is constitutively transcribed into a (+) strand RNA molecule from a RNA polymerase II promoter in the nucleus of the host cell, but little or no expression of the reporter gene occurs until the cell is infected by a positive-strand RNA virus that recognizes the *cis*-acting sequences and causes significant replication of the (+) strand RNA molecule through a (-) strand RNA intermediate.

Recently, a review article on the use of transgenic cell lines for detecting animal viruses speculated that a similar strategy could be used to make a cell line for detecting negative-strand RNA viruses (Olivo et al., *supra*). The article stated generally that such a cell line would constitutively express the viral nucleocapsid protein and a chimeric antigenomic RNA molecule which contains a reporter gene open reading frame (ORF) and the *cis*-acting sequences necessary for replication and transcription by the replicase and transcriptase of the same negative-strand RNA virus. This chimeric RNA molecule would be designed, in an unspecified manner, to preclude translation of the reporter gene in uninfected cells. Replication and transcription of the chimeric RNA molecule and synthesis of translatable reporter gene mRNA would be carried out by the replicase-transcriptase complex brought into this hypothetical cell by infection with the negative-strand RNA virus. However, this article did not teach how the nucleocapsid protein and chimeric RNA molecule would be constitutively expressed or how to design the chimeric RNA molecule to preclude translation of the reporter gene in the absence of infectious virus. It would be desirable, therefore, to provide a rapid, specific, sensitive and cost-efficient assay for the detection of infectious negative-stranded RNA viruses.

Summary of the Invention

Briefly, therefore, the present invention is directed to novel compositions and methods which permit the detection of a negative-strand RNA virus in a specimen. In one embodiment, the invention provides a genetically engineered cell which expresses a reporter gene product only in the presence of infectious virions of the negative-strand RNA virus. The genetically engineered cell comprises a polynucleotide encoding a DNA-dependent RNA polymerase and a cDNA comprising a minigenome and a miniantigenome of the negative-strand RNA virus operably linked to a promoter for the DNA-dependent RNA polymerase. The miniantigenome comprises a

nucleotide sequence encoding a reporter gene product whose expression is dependent upon the presence of the negative-strand RNA virus.

Expression of the DNA-dependent RNA polymerase in the cell results in transcription of the cDNA into a minigenome RNA or a miniantigenome RNA, depending on the orientation of the promoter with respect to the open reading frame (ORF) of the reporter gene. When the promoter is located downstream of the ORF, a minigenome RNA is synthesized which contains an untranslatable negative-sense copy of the reporter gene ORF and when the promoter is located upstream of the ORF, a miniantigenome RNA is synthesized. Although this miniantigenome contains a positive-sense copy of the reporter gene ORF, its translation is very inefficient because the miniantigenome is not a capped, polyadenylated mRNA. Thus, detectable expression of the reporter gene in the absence of infectious virus does not occur. The cell also comprises one or more nucleotide sequences encoding each of the nucleocapsid proteins of the negative-strand RNA virus which are necessary and sufficient for replication of the minigenome RNA or miniantigenome RNA synthesized by the DNA-dependent RNA polymerase. Minigenome RNA synthesized in the presence of these nucleocapsid proteins, which includes minigenome RNA made as an intermediate during replication of the miniantigenome, is stabilized and amplified without detectable expression of the reporter gene product. Infection of the cell with the negative-strand virus introduces all the viral gene products necessary for transcription of the preformed minigenome RNA to produce translatable mRNA molecules encoding the reporter gene product.

In another embodiment, the invention provides a method for detecting a negative-strand RNA virus in a sample which comprises culturing the above-described genetically engineered cell for a time sufficient to synthesize the minigenome or miniantigenome RNA and the nucleocapsid proteins, then incubating the cells with the sample and detecting expression of the reporter gene product. In some embodiments, the method includes quantifying the number of infectious particles of the RNA virus that are present in the sample.

In other embodiments the genetically engineered cell is used to screen compounds for anti-viral activity and for measuring antibody responses of individuals infected with or vaccinated against the RNA virus.

In yet another embodiment, the invention provides a kit for detecting the presence of a negative-strand RNA virus in a sample which comprises a supply of the above-described genetically engineered cells. In a preferred embodiment, the kit also includes a set of reagents for detecting expression of the reporter gene product.

5 The invention also provides a method for making a genetically-engineered cell capable of detecting a negative-strand RNA virus. The method comprises (1) providing a cell that is susceptible to infection by the negative-strand RNA virus; (2) introducing into the cell a polynucleotide encoding a DNA-dependent RNA polymerase and selecting for cells containing the polynucleotide; (3) introducing into
10 the selected cells a cDNA comprising a minigenome and an miniantigenome of the negative-strand RNA virus operably linked to a promoter for the DNA-dependent RNA polymerase, wherein the miniantigenome comprises a nucleotide sequence encoding a reporter gene product, and wherein expression of the reporter gene product is dependent upon the presence of the negative-strand RNA virus; and (3) introducing
15 into the cells one or more nucleotide sequences encoding each of the nucleocapsid proteins of the negative-strand RNA virus which are necessary and sufficient for replication of minigenome RNA or miniantigenome RNA synthesized by the DNA-dependent RNA polymerase.

 Among the several advantages of the present invention may be noted the
20 provision of a genetically engineered cell which expresses a reporter gene product only after infection by a particular negative-strand RNA virus; the provision of a rapid and sensitive assay for specifically detecting the negative-strand RNA virus; the provision of such an assay that is capable of quantitating the amount of the negative-strand RNA virus in a sample that can be useful for assessing compounds for antiviral
25 activity and for measuring antibody responses in infected or vaccinated individuals; and the provision of a kit for detecting and/or quantitating the negative-strand RNA virus.

Brief Description of the Drawings

30 Figure 1A is a schematic representation of SR19/pac cDNA, showing an SP6 promoter operably linked to a DNA copy of a noncytopathic Sindbis virus replicon in which the gene for puromycin acetyltransferase (PAC) is operably linked to the Sindbis subgenomic promoter;

Figure 1B is a schematic representation of SR19/T7pol/pac cDNA, which is identical to the cDNA of Figure 1A except that the noncytopathic Sindbis virus replicon contains a the T7 RNA polymerase operably linked to a second Sindbis subgenomic promoter;

Figure 2 is a bar graph of β -galactosidase activity in baby hamster kidney (BHK) cells transformed with a noncytopathic Sindbis virus replicon expressing T7 RNA polymerase (SR19/T7/pac) or the parent replicon (SR19/pac) and transfected with a T7 expression plasmid expressing the *E. coli lacZ* gene under the control of the T7 promoter (pTM1-LacZ) or the parent T7 expression plasmid (pTM1);

Figure 3A is a bar graph of β -galactosidase activity in mock-infected or RSV-infected SR19/T7/pac-BHK or SR19/pac-BHK cells transfected with a cDNA of a *lacZ*-containing RSV minigenome (pMP210LacZ) alone or with T7 expression plasmids expressing the RSV nucleocapsid proteins N, L and P (+ NLP);

Figure 3B is a bar graph of β -galactosidase activity in mock-infected or RSV-infected SR19/T7/pac-BHK or SR19/pac-BHK cells transfected with pMP210LacZ alone or with various combinations of the T7 expression plasmids for N, L and P as indicated by the + and - symbols below the graph;

Figure 4A is a graph of β -galactosidase activity in SR19/T7/pac-BHK cells cotransfected with pMP210LacZ and the nucleocapsid expression plasmids and infected with the indicated amounts of RSV;

Figure 4B is a graph of β -galactosidase activity in the cells from Figure 4A at the indicated times following RSV infection at a high multiplicity of infection (HMOI) or at a low multiplicity of infection (LMOI);

Figure 5 illustrates the effect of ribivarin on the induction of reporter gene activity in RSV-infected SR19/T7/pac-BHK cells cotransfected with pMP210LacZ and the nucleocapsid expression plasmids showing in FIG. 5A a bar graph of β -galactosidase activity at high concentrations of ribivarin and in FIG. 5B a dose-response curve of β -galactosidase activity at subinhibitory concentrations of ribivarin expressed as a percentage of the control;

Figure 6 is a graph of β -galactosidase activity in SR19/T7/pac-BHK cells cotransfected with pMP210LacZ and the nucleocapsid expression plasmids after infection with RSV which had been preincubated with the indicated dilutions of pooled human immune globulin; and

Figure 7 is a schematic representation of the recombinant constructs and molecular pathways involved in generating a reporter gene product in accordance with one embodiment of the invention.

5 Description of Preferred Embodiments

In accordance with the present invention, a method for detecting negative-strand RNA viruses and genetically engineered cells for use in the method are provided. The invention is based on the inventors' discoveries (1) that minigenome RNA of a negative-strand virus expressed by a genetically engineered cell in the
10 presence of certain viral nucleocapsid proteins is accessible for transcription of translatable reporter gene mRNA when the cell is subsequently infected with the negative-strand virus and (2) that expression of the product of the reporter gene is not detectably expressed in the absence of the negative-strand virus.

In the context of this disclosure, the following terms shall be defined as
15 follows unless otherwise indicated:

"cis-acting sequences" means the nucleotide sequences from an RNA virus genome that are necessary for the recognition of the genomic RNA by specific protein(s) of the RNA virus that carry out replication and/or transcription of the genome;

20 "heterologous DNA-dependent RNA polymerase" means a DNA-dependent RNA polymerase not naturally encoded in a vertebrate cell;

"infectious" when used to describe a virus or an RNA molecule, means a virus or RNA molecule that is self-replicating and provides for transcription of viral mRNAs in a host cell;

25 "miniantigenome" means an RNA molecule or a DNA molecule that is complementary to a minigenome;

"minigenome" means a DNA or RNA analog of the genome of a negative-strand RNA virus which contains the cis-acting sequences necessary for replication and transcription in cells infected with the negative-strand RNA virus and which
30 contains a negative-sense open reading frame of a reporter gene in place of one or more of the viral protein-coding sequences;

"negative-strand RNA virus" means a virus whose genome consists of one negative-sense RNA molecule (nonsegmented genome) or a plurality of RNA

molecules of negative sense (segmented genome) and which replicates its genome through a positive-sense RNA intermediate(s);

"replicon" means a replication-competent viral RNA that contains the genetic information needed for virus replication but not for virus assembly;

5 "transfected cell" or "transformed cell" means a cell containing an exogenously introduced nucleic acid molecule which may be present in the nucleus or the cytoplasm of the cell;

"stably transformed cell" means a cell containing an exogenously introduced nucleic acid molecule which is present in the nucleus of the cell and may be stably
10 integrated into the chromosomal DNA of the cell.

A genetically engineered cell according to the present invention is a vertebrate cell that is susceptible to infection by the negative-strand RNA virus to be detected and that is capable of expressing a heterologous DNA-dependent RNA polymerase which synthesizes a minigenome RNA or miniantigenome RNA of a negative-strand
15 virus from a promoter recognized by the DNA-dependent RNA polymerase. The cell is also capable of expressing the nucleocapsid proteins of the negative-strand virus that are necessary for replication of the minigenome RNA or miniantigenome RNA.

A cell is susceptible to infection if the infecting virus can enter the cell and proceed far enough in its replication cycle to express proteins necessary for efficient
20 synthesis of translatable reporter gene mRNA from preformed minigenome RNA. It will be understood by those skilled in the art that cell lines susceptible to infection by a particular negative-strand RNA virus can be readily identified by searching the literature for known susceptible cell lines and/or by screening candidate cell lines using well-known procedures requiring only routine experimentation. Susceptible
25 cells for negative-strand RNA viruses include, but are not limited to baby hamster kidney cells, African green monkey cells, rabbit skin fibroblasts, 3T3 mouse cells and the like. Baby hamster kidney cells are preferred for use as the host cell for detecting negative-strand viruses that infect mammals, including humans.

The heterologous DNA-dependent RNA polymerase expressed by the cell comprises
30 a naturally-occurring amino acid sequence or a functional equivalent thereof. A functional equivalent of a DNA-dependent RNA polymerase contains one or more amino acid substitutions, additions or deletions in the naturally-occurring amino acid sequence that do not prevent the modified polymerase from synthesizing full-length transcripts from a DNA

template. Typically, the heterologous DNA-dependent RNA polymerase comprises the amino acid sequence of an RNA polymerase made by a bacteriophage such as T7, T3 or SP6. A preferred DNA-dependent RNA polymerase for use in the present invention is T7 RNA polymerase.

5 In one embodiment, the heterologous DNA-dependent RNA polymerase is encoded by a polynucleotide integrated into the cell nucleus and comprises a polymerase II promoter recognized by the host cell transcription machinery operably linked to a nucleotide sequence encoding the heterologous DNA-dependent RNA polymerase. In a preferred embodiment, the polynucleotide encoding the

10 heterologous DNA-dependent RNA polymerase is an RNA molecule that comprises a noncytopathic positive-strand virus replicon. Use of a noncytopathic replicon allows for long-term, continuous expression of the DNA-dependent RNA polymerase without killing the host cell. Modification of replicons to make them noncytopathic is known in the art and can be accomplished, i.e., by isolating variants having adaptive

15 mutations in the viral replication machinery that allows persistent noncytopathic replication in vertebrate cells or by using a dominant selectable marker (see e.g., Frolov et al., *Proc. Natl. Acad. Sci. USA* 93:11371-11377, 1996 and referenced cited therein). Preferred viruses for preparation of noncytopathic replicons include flaviviruses, e.g., Kunjin virus, alphaviruses such as Sindbis virus, Semliki forest

20 virus, nodaviruses such as flock house virus, astroviruses and coronaviruses. The noncytopathic replicon can be transcribed from a cDNA of the replicon that is operably linked to a polymerase II promoter and integrated into the nucleus. Alternatively, the replicon can be prepared chemically or by *in vitro* transcription from a plasmid and then transfected into the cell cytoplasm.

25 The heterologous DNA-dependent RNA polymerase specifically recognizes a promoter in a cDNA comprising a minigenome and a miniantigenome of the negative-strand RNA virus to be detected. Typically, the nucleotide sequence of the promoter is identical to a promoter sequence found in the same organism that encodes the DNA-dependent RNA polymerase. However, the promoter may contain one or more

30 nucleotide substitutions in the naturally-occurring sequence, as long as such substitutions do not affect specificity of recognition by the DNA-dependent RNA polymerase. The promoter is operably linked to either the DNA sequence encoding the minigenome or to the DNA sequence encoding the miniantigenome to direct the

synthesis of either minigenome RNA or miniantigenome RNA, respectively.

Preferably, the promoter directs the synthesis of minigenome RNA.

The minigenome encoded by the cDNA comprises the cis-acting 3'-leader and 5'-trailer sequences from the viral genome that are necessary for its replication and transcription by the viral-encoded RNA-dependent RNA polymerase. For the detection of RNA viruses having segmented genomes, it is believed that cis-acting sequences from any of the segments will make a functional minigenome. These cis-acting sequences flank a negative-sense copy of a reporter gene ORF, which is operably linked to gene-start (GS) and gene-end/polyadenylation (GE) transcription signals that are recognized by the RNA-dependent RNA polymerase. The leader, trailer, GS and GE sequences for any particular negative-strand RNA virus can be found in the scientific literature or can be identified by the skilled artisan using well-known and routine techniques. See, e.g., Weng et al., *Genome Res.* 5:202-207, 1995; Garcia-Sastre et al., *Annu. Rev. Microbiol.* 47:765-790, 1993 and references cited therein. In addition, it is intended that the minigenome can contain modifications in the naturally-occurring leader, trailer, GS and GE sequences that do not destroy replication or transcription of the minigenome or production of reporter gene mRNA in the presence of the negative-strand virus.

Any reporter gene that encodes a detectable product is suitable for use in the present invention. The reporter gene product is preferably one that can easily be assayed for or detected in a cell. One enzyme that has proved to be particularly useful as a reporter gene product is β -galactosidase. Preferably, a bacterial β -galactosidase is used, and most preferably the β -galactosidase from *E. coli* that is encoded by the LacZ gene. β -galactosidase is preferred because of its well-characterized nature and the existence of a variety of methods to detect its presence. Other reporter gene products useful in this invention generally include hydrolases or oxidoreductases and, in particular, such enzymes as β -glucosidase, β -glucuronidase, β -hexosaminidase, luciferase, phospholipase, phosphatase, etc. Green fluorescent protein (GFP) is another reporter gene product useful in the invention.

An ORF encoding β -galactosidase or luciferase is particularly preferred for use in this invention because of the numerous methods known to detect expression of these enzymes and the relative sensitivity of such methods. Among these methods include histochemical assays involving a chromogenic or fluorogenic substrate which

permits detection of β -galactosidase activity by a change in the color of the cell that can be detected macroscopically or microscopically. The use of luciferase provides an enzymatic assay that is more sensitive than the colorimetric or fluorometric β -galactosidase assay. Expression of luciferase may be detected by known

5 luminometric methods using luciferin as the enzyme substrate.

The minigenome or miniantigenome synthesized by the DNA-dependent RNA polymerase should have replication competent termini. These can be generated by appropriate construction of the minigenome-encoding cDNA using methods known in the art. For example, the cDNA can include at least one transcription terminator
10 recognized by the DNA-dependent RNA polymerase in operable linkage with the minigenome or the miniantigenome. More preferably, the cDNA also comprises a self-cleaving ribozyme located between the transcription terminator and the 3'-terminus of the minigenome or miniantigenome. The position of the ribozyme is such that a transcript of the cDNA synthesized by the DNA-dependent RNA polymerase is
15 cleaved by the ribozyme to produce a minigenome RNA or miniantigenome RNA having a replication competent 3'-terminus. Similarly, the cDNA can also contain a self-cleaving ribozyme positioned between the promoter for the DNA-dependent RNA polymerase and the trailer or leader sequence of the minigenome or miniantigenome, respectively, such that the 5'-terminus is replication competent. A
20 number of self-cleaving ribozymes suitable for use in the invention are known in the art. See, e.g., Grosfeld et al., *supra*; Long et al., *FASEB J.* 7:25-30, 1993; Perrotta et al., *Nature* 350:434-436, 1991.

The genetically engineered cell also comprises one or more nucleotide sequences encoding the nucleocapsid proteins that are necessary and sufficient for
25 replication of the minigenome RNA or miniantigenome RNA transcribed from the cDNA. As used herein, "necessary and sufficient for replication" is intended to mean that minigenome RNA or miniantigenome RNA synthesized from the cDNA in the presence of these nucleocapsid proteins is replicated in the absence of the negative-strand virus to be detected while detectable expression of the reporter gene product
30 does not occur until the cell is infected with the negative-strand virus. As used herein, the terms "detectable expression of the reporter gene product" and "dependent upon the presence of the negative-strand virus" mean that the reporter gene product is practically detectable above background levels in the particular diagnostic assay being

used, with background levels referring to measurements taken when the assay is performed in the absence of infectious virus or with genetically-engineered cells that lack the reporter gene.

As demonstrated below, the nucleocapsid proteins necessary and sufficient for replication of an RSV minigenome are the N, P and L proteins. Functional counterparts of these proteins in other negative-strand viruses can be identified from the scientific literature and by using methods known in the art. The combination of nucleocapsid proteins that are necessary and sufficient to replicate the minigenome of any particular negative-strand virus can be determined as described below in Example 3. In brief, T7-expression plasmids are constructed that express each of the nucleocapsid proteins and various combinations of these plasmids are introduced into a vertebrate cell expressing T7 RNA polymerase and containing a minigenome-encoding cDNA. The resulting genetically-engineered cell is incubated for a time sufficient for minigenome replication to proceed and then the cell is mock-infected or infected with the negative-strand virus and expression of the reporter gene product is measured. A combination of nucleocapsid proteins suitable for use in the present invention is one whose pre-infection expression results in cells that lack detectable reporter gene product in the absence of virus but that show efficient expression of the reporter gene product in the presence of the virus. It is intended that any or all of these nucleocapsid proteins may contain amino acid substitutions or deletions in the naturally-occurring sequence as long as the modified protein(s) functions in an equivalent manner as the native protein.

The nucleotide sequences encoding the desired nucleocapsid proteins can be provided by separate polynucleotides cotransfected into the cell or can be provided by a single polynucleotide in which separate transcription units encode the desired nucleocapsid proteins. Expression of mRNAs for the nucleocapsid proteins can be directed by promoters and transcription termination signals recognized by the heterologous DNA-dependent RNA polymerase expressed in the host cell cytoplasm or by polymerase II in the host cell nucleus. In embodiments in which a noncytopathic replicon expresses the DNA-dependent RNA polymerase, the replicon can also contain the nucleotide sequences encoding the nucleocapsid proteins arranged such that the replicon expresses mRNAs for each of the nucleocapsid proteins from a different subgenomic promoter. In a preferred embodiment, the

heterologous DNA-dependent RNA polymerase is T7, T3, or SP6 RNA polymerase and the nucleotide sequences for the desired nucleocapsid proteins are operably linked to the corresponding T7, T3, or SP6 promoter.

In accordance with the method provided by the invention, the above-described
5 genetically-engineered cells can be used to test a sample for the presence of a negative-strand virus. The sample can be any material which can be placed into a fluid or fluid environment and includes biological fluids such as blood, semen, nasopharyngeal swabs, cerebrospinal fluids and the like.

To carry out the method, the genetically engineered cells are first cultured for
10 a period of time sufficient for synthesis of the minigenome or miniantigenome RNA and for expression of the nucleocapsid proteins. It is believed this culture step provides the minigenome RNA in an encapsidated form that is accessible for efficient expression of the reporter gene product by the replicase/transcriptase machinery of the incoming virus. Typically, this first culture period will be between 16 and 24 hours,
15 but shorter or longer culture times may also produce functional, encapsidated minigenome RNA and the culture time suitable for a particular assay can be determined empirically.

The pre-cultured cells are then incubated with the sample in a suitable culture medium for a period of time sufficient for the infectious cycle of the target RNA virus
20 to proceed. If the target virus is in the specimen, it will produce the factor(s) necessary for detectable expression of the reporter gene product. Typically, this incubation period is between 24 and 48 hours, although the optimal length for any particular negative-strand RNA virus should be determined empirically. Expression of the reporter gene product can be detected in the cells, in the culture medium, or in
25 cell extracts.

The invention also provides a kit for detecting a negative-strand RNA virus. The kit is prepared by placing in a container a supply of the above-described genetically-engineered cells sufficient to conduct an assay or a number of assays in accordance with the invention. Preferably, the kit is also provided with reagents
30 necessary for detecting the reporter gene product, placed in separate containers. An instruction manual can also be included in the kit.

The method and cells of this invention are useful for the detection of negative-strand RNA viruses belonging to the following families: *Rhabdoviridae*, *Filoviridae*, *Paramyxoviridae*, *Orthomyxoviridae*, *Bunyaviridae*, and *Arenaviridae*.

The following examples of the present invention are offered by way of illustration
5 and are not to be considered in a limiting sense.

Example 1

This example illustrates that T7 RNA polymerase expressed by a Sindbis virus replicon in baby hamster kidney (BHK) cells is functional.

10 As discussed above, studies of replication and transcription in negative-strand viruses have primarily used a vaccinia virus/T7 RNA polymerase system to supply intracellular T7 RNA polymerase for transcription of a desired gene under the control of the T7 promoter. However, an RNA detection method requiring coinfection with vaccinia virus would not be practicable for clinical diagnostic purposes. Thus, the inventors herein wished to develop a
15 system that would permit the use of stably transformed cells, which would be easier to handle and allow more widespread use.

Recently, noncytopathic Sindbis virus replicons have been developed which contain the gene for puromycin resistance (*pac*) under control of the Sindbis subgenomic RNA promoter (SR19/*pac*) (Frolov et al., *Proc. Natl. Acad. Sci. USA* 93:11371-11377, 1996).
20 Transfection of BHK cells with RNA transcribed *in vitro* from a cDNA of an SR19/*pac* replicon (Fig. 1A) produces a population of puromycin-resistant cells due to constitutive replication and transcription of the noncytopathic replicon and consequent expression of puromycin acetyl transferase. Modification of SR19/*pac* replicons to express an additional foreign gene operably linked to a second subgenomic RNA promoter has been achieved
25 (Lindenbach et al., *J. Virol.* 71:9608-9617, 1997). For example, cells that carry a replicon expressing T7 RNA polymerase (SR19/T7Pol/*pac*) can transcribe a gene under the control of the T7 promoter following transfection of that gene (Agapov, E., et al., *Proc. Natl. Acad. Sci. USA* 95:12989-12994, 1998).

To determine if the SR19/T7pol/*pac* system could be used to express functional T7
30 RNA polymerase in BHK cells, replicon RNAs were transcribed *in vitro* from SR19/*pac* and SR19/T7Pol/*pac* cDNA plasmids (Figs. 1A, 1B) (gift of Eugene Agapov and Charles Rice, Washington University School of Medicine, St. Louis, MO) using SP6 RNA polymerase. The replicon RNAs were electroporated into BHK-21 cells (ATCC) and replicon-containing cell populations were selected for puromycin resistance as described by Lindenbach et al.,
35 *supra*. The SR19/*pac*- and SR19/T7Pol/*pac*-transformed cells were then transfected with the

T7 expression vector pTM1 (a generous gift of Bernard Moss, NIAID, Bethesda, MD) or with pTM1-LacZ, which was generated by cloning the *E. coli lacZ* gene into the *NcoI-BamHI* window of the pTM-1 plasmid, thus placing the *lacZ* gene under control of the T7 promoter. The four groups of recombinant cells were maintained in puromycin-containing medium for 24 hours and then β -galactosidase activity in lysates prepared from these cells was determined as follows.

β -galactosidase activity was measured using the substrate chlorophenolred- β -D-galactopyranoside (CPRG, Boehringer Mannheim, Indianapolis, IN) (final concentration 5 mM) in a 0.2 M potassium phosphate buffer, pH 7.8, with 1 mM $MgCl_2$. Lysates were made in this buffer containing 1% Triton X-100 and 1 mM dithiothreitol. Fifty μ liters of lysate were mixed with 50 μ l of substrate in a microtiter plate well. After incubation for 30 to 120 min at room temperature, the optical density at a wavelength of 562 nm (OD_{562}) was measured with a THERMOmax microplate reader using SOFTmax software (Molecular Devices, Sunnyvale, CA). The assay was shown to be linear up to an OD_{562} of 3.0.

The amount of β -galactosidase activity in the four different groups of cells is shown in Fig. 2 (amounts shown are the mean of duplicate samples). High levels of β -galactosidase activity were detected in cells containing both the SR19/T7Pol/pac replicon and the pTM1-LacZ plasmid, but not in control cells that lacked the T7 RNA polymerase or the *lacZ* gene (Figure 2). Thus, functional T7 RNA polymerase is expressed in BHK cells transfected with SR19/T7pol/pac RNA.

Example 2

This example illustrates that SR19/T7Pol/pac-transformed BHK cells are susceptible to infection by the negative-strand virus RSV and that such RSV-infected cells support replication and transcription of an RSV minigenome.

SR19/T7Pol/pac BHK cells were mock-infected or infected with 10 plaque-forming units (pfu)/cell of RSV (strain A2), which was propagated and titered on HEp-2 cells as described previously (Grosfeld et al., *supra*). Forty-five minutes later, the cells were transfected over a 3-h period with 0.5-1.5 μ g of *in vitro*-synthesized RSV-CAT minigenome RNA, which contains the bacterial *cat* gene in place of all of the viral protein open reading frames (Collins et al, *Proc. Natl. Acad. Sci. USA* 88:9663-9667, 1991). The minigenome RNA was a run-off transcript of 935 nt produced by T7 RNA polymerase of *HgaI*-linearized RSV-CAT cDNA (Collins et al., 1991). Only RSV-infected cells exhibited reporter gene

expression (data not shown), indicating that SR19/T7Pol/pac BHK cells are permissive for RSV infection as well as RSV minigenome replication and expression.

The inventors next asked whether minigenome RNA transcribed *in vivo* in SR19/T7Pol/pac BHK cells was functional. SR19/T7Pol/pac BHK cells were mock-infected or infected with RSV (strain A2) at 10 plaque-forming units (pfu) per cell and 45 min. later were transfected with pC2CAT, a derivative of RSV-CAT cDNA which, *inter alia*, contains a hammerhead-type ribozyme sequence and T7 transcription terminator at the 3' end of the encoded transcript (Grosfeld et al., 1995, *supra*). Ribozyme self-cleavage of the T7-synthesized transcript of pC2CAT leaves a single non-RSV-specific 3'-phosphorylated U residue on the 3' terminus (Grosfeld et al., 1995, *supra*). Under these conditions, RSV-infected cells exhibited a marked increase in CAT activity as compared to CAT activity in mock-infected cells (data not shown).

The results of these experiments indicate that RSV-infected SR19/T7Pol/pac cells can produce a functional RSV minigenome from minigenome RNA or minigenome cDNA that is subsequently transfected into the cell.

Example 3

This example illustrates that preformed minigenome RNA requires RSV nucleocapsid proteins for activity after RSV infection.

The assay initially envisioned by the inventors for detecting RSV in cell culture involved constitutive expression of an RSV minigenome RNA that would be replicated and transcribed after subsequent infection with RSV. To test the feasibility of this protocol, the following experiment was performed.

SR19/T7Pol/pac cells were transfected with pMP210, a cDNA of an RSV minigenome containing a negative-sense copy of the gene for green fluorescent protein (GFP), and 6-24 h later the cells were infected with 10 pfu of RSV. pMP210 was made from pC2Luc cDNA, which contains the luciferase gene substituted for the CAT gene in the above-described pC2CAT construct (Collins et al., *Proc. Natl. Acad. Sci. USA* 93:81-85, 1996). A PCR product containing the Green Lantern Protein gene (Life Technologies, Gaithersburg, MD) flanked by gene start (GS) and gene end (GE) signals was inserted into the BstXI site in the leader of pC2Luc, and the luciferase gene was deleted. In addition, a 5' hammerhead ribozyme preceded by a unique *AvrII* site (Altschuler et al., *Gene* 122:85-90, 1992) was inserted just before the trailer sequence. The pC2 3' hammerhead ribozyme was replaced with a PCR product containing the antigenomic hepatitis delta virus ribozyme (Perrotta et al., *Nature* 350:434-436, 1991) followed by a unique *XhoI* site.

Under these conditions, no RSV-dependent GFP expression was observed. These negative results suggest that minigenome RNA has a very short half-life in uninfected BHK cells or is in some manner not accessible to or recognized by the incoming RSV replicase and/or transcriptase. Thus, a viable assay for detecting RSV infection using an RSV minigenome requires a strategy for stabilizing the minigenome RNA and/or making it replication competent.

It has been shown that RSV N, P, and L proteins are necessary and sufficient for replication of RSV minigenomes and that expression of the N, P, and L proteins leads to transcription of RSV mRNA, although not necessarily full-length mRNA (Grosfeld et al., *supra*; Yu et al., *supra*. In addition, expression of only the N and P proteins leads to encapsidation of RNA minigenomes and this is increased 10 to 50-fold by the addition of the L protein (Atreya et al., *J. Virol.* 72:1452-1461, 1998). Based on these results, the inventors investigated whether a minigenome expressed in the presence of the N, P, and L proteins would be accessible to transcription of reporter gene mRNA when the cells were subsequently infected with RSV.

SR19/T7Pol/pac cells or SR19/pac cells were transfected with 1 µg of pMP210LacZ, a *lacZ*-containing minigenome cDNA, with or without 0.2 µg each of the T7 expression plasmids pTM1-N, pTM1-P, and pTM1-L described in Grosfeld et al., *supra*. pMP210LacZ was made by replacing the ORF of *GFP* in pMP210 with the ORF of *lacZ* using the unique *Xba*I and *Xma*I sites. Twenty hours after transfection the cells were trypsinized and plated in 24 well dishes and six hours later infected with RSV (5 pfu/cell) or mock-infected. At 36 hrs after infection, the cells were lysed and assayed for β-galactosidase as described above. The results, which are the mean of duplicate samples, are shown in Figure 3A.

Significant induction of β-galactosidase activity by RSV infection was observed in cells expressing both T7 RNA polymerase and the RSV nucleocapsid proteins N, L and P. These cells are illustrated in Figure 7. Notably, β-galactosidase activity in the corresponding mock-infected cells was not significantly higher than β-galactosidase activity in mock-infected cells lacking T7 (background) or in mock-infected cells lacking N, L and P.

To assess the relative requirement for each of the nucleocapsid proteins, various combinations of the N, L and P expression plasmids (0.2 µg each) were cotransfected with the minigenome cDNA (pMP210LacZ at 1 µg) into SR19/T7Pol/pac cells and the assay was carried out as above. The results, which are the mean of triplicate samples ± the standard deviation, are shown in Fig. 3B. RSV-induced β-galactosidase activity was significant only in the presence of all three nucleocapsid proteins. In addition, the amount of activity was unchanged when the assay was carried out after 10 passages of these minigenome-expressing cells.

When evaluated by histochemical staining as described in U.S. Patent 5,418,132, β -galactosidase-positive infected cells were observed only in the presence of the nucleocapsid proteins: a few stained infected cells per well were observed with N alone or with N plus P, while many stained infected cells were consistently observed when N, P, and L were all expressed (data not shown). Significantly, no stained cells were observed in the absence of RSV with or without the nucleocapsid proteins. The requirement for all three nucleocapsid proteins in this assay suggests that encapsidation and replication of the minigenome RNA are necessary for efficient reporter gene expression by the incoming RSV. In addition, the lack of β -galactosidase activity in mock-infected cells expressing all three nucleocapsid proteins indicates that detectable expression of β -galactosidase is dependent upon some factor(s) provided by RSV infection, probably the M2-1 protein.

Example 4

This example illustrates that the level of expression of the minigenome reporter gene is dependent on the input concentration of RSV.

Approximately 2×10^6 SR19/T7Pol/pac cells in six 9.5 cm² wells were transfected with pMP210LacZ (1 μ g) and pTM1-N, P, and L (0.2 μ g each). After 18 h the transfected cells were trypsinized, pooled and plated into all the wells of a 48-well plate. Six h later approximately 10^5 cells were either (A) infected with increasing volumes of a two-fold dilution of a stock RSV (titer 2×10^7 pfu/ml assayed on HEp2 cells) and β -galactosidase activity assayed 36 hr later or (B) infected with either 0.5 microliters (low multiplicity of infection; LMOI) or 25 microliters (high multiplicity of infection; HMOI) of the same stock virus and β -galactosidase activity assayed at the indicated times after infection. The results of the dose response assay are shown in Fig. 4A (expressed as the mean of triplicate samples \pm the standard deviation) and the results of the time course assay are shown in Fig. 4B (the mean of duplicate samples).

The assay exhibited a relatively linear dose-response for RSV over a range of greater than one log₁₀ (Figure 4A). The RSV-induced expression exhibited a plateau effect above an inoculum equivalent to a multiplicity of infection of approximately 0.6 pfu/cell (based on HEp2 cells). These results indicate that this assay could be used in place of a plaque assay to quantitate RSV stocks.

When β -galactosidase in these minigenome-expressing cells was measured as a function of time after infection, β -galactosidase activity following a LMOI infection (0.2 pfu/cell) reached the same level as that following a HMOI infection (10 pfu/cell), but with a

delay of approximately 48 h (Fig. 4B). This suggests that RSV replicates and spreads within these cell cultures and that the duration of an infectious cycle is approximately 48 h.

Example 5

5 This example illustrates that RSV-induced expression of the minigenome reporter gene is sensitive to ribavirin.

Ribavirin inhibits *in vitro* replication of many RNA viruses including RSV (Sidwell et al., *Pharmacol Ther* 6:123-146, 1979). Depending on the assay used, the 50% effective dose (ED₅₀) of ribavirin for RSV has been reported to be around 5 µg/ml (Chiba et al., *Biol Pharm Bull* 18:1081-1083, 1995). To determine if ribavirin affects the detection of RSV by a minigenome assay, SR19/T7Pol/pac cells were cotransfected with pMP210LacZ, pTM1-N, pTM1-P, and pTM1-L as described in Example 3 and then infected with RSV (5 pfu/cell) in the presence of 1, 5, 50 or 200 µg/ml of ribavirin. As shown in Fig. 5A, β-galactosidase activity was reduced to near background levels by as little as 1 µg/ml ribavirin (Fig. 5A).
15 When this dose response experiment was repeated at subinhibitory concentrations between 0 and 400 ng/ml ribavirin, the ED₅₀ of ribavirin was determined to be less than 100 ng/ml (Fig. 5B).

Treatment of the cells with ribavirin (1 µg/ml) from the time of transfection of the RSV cDNA plasmids until the time of RSV infection had no effect on RSV-dependent β-galactosidase expression (data not shown). This suggests that under the conditions in which this assay was performed, ribavirin primarily affects RSV replication and transcription rather than the Sindbis replicon or T7 polymerase transcription and than any minigenome replication mediated by the RSV L protein occurring prior to replication of incoming RSV must be relatively insensitive to ribavirin.

25 Example 6

This example illustrates that RSV-induced β-galactosidase activity in minigenome-expressing cells is sensitive to neutralizing antibody to RSV.

To demonstrate that the RSV-induced β-galactosidase activity in this system requires infectious virus, and can, therefore, be blocked by neutralizing antibody, we incubated RSV with different dilutions of a sample of pooled human gamma globulin. RSV is a highly prevalent human pathogen and most adults have been infected at least once (Collins et al., Respiratory Syncytial Virus, pp. 1313-1350 in *Fields Virology*, B. N. Fields, D. M. Knipe and P. M. Howley, eds., Lippencott-Raven, Philadelphia, 1996). Therefore, pooled human gamma globulin has significant neutralizing activity against RSV. The globulin-treated virus
35 was used to infect SR19/T7Pol/pac cells that had previously been transfected with

pMP210LacZ and pTM1-N, pTM1-P, and pTM1-L. As shown in Figure 6, β -galactosidase activity was inversely proportional to the amount of dilution of the gamma globulin sample.

Example 7

5 This example illustrates the specificity of RSV minigenome-expressing cells in detecting RSV in patient samples.

 SR19/T7Pol/pac cells that had previously been transfected with pMP210LacZ and pTM1-N, pTM1-P, and pTM1-L were incubated with two patient specimens, previously determined to contain RSV, or with patient specimens known to contain influenza A,
10 influenza B, and parainfluenza types 1 and 2. Induction of β -galactosidase activity was determined by histochemical staining.

 Incubation with the RSV-containing specimens produced significant β -galactosidase, while specimens containing the other negative-stranded viruses did not induce any positively stained cells (data not shown). Thus, the assay is specific for RSV.

15 The sensitivity of this assay can be enhanced by increasing the time of infection before assaying for β -galactosidase activity. At 36 h after infection, 3000 pfu could be detected; at 5 days after infection even an inoculum of less than 10 pfu gave a reading several-fold above background.

 In view of the above, it will be seen that the several advantages of the
20 invention are achieved and other advantageous results attained.

 As various changes could be made in the above methods and compositions without departing from the scope of the invention, it is intended that all matter contained in the above description and shown in the accompanying drawings shall be interpreted as illustrative and not in a limiting sense.

25 All references cited in this specification, including patents and patent applications, are hereby incorporated by reference. The discussion of references herein is intended merely to summarize the assertions made by their authors and no admission is made that any reference constitutes prior art. Applicants reserve the right to challenge the accuracy and pertinency of the cited references.

What is Claimed is:

1. A genetically engineered cell for detecting a negative-strand RNA virus, the cell comprising:
 - a polynucleotide encoding a DNA-dependent RNA polymerase;
 - a cDNA comprising a minigenome and a miniantigenome of the negative-strand
 - 5 RNA virus operably linked to a promoter for the DNA-dependent RNA polymerase, wherein the miniantigenome comprises a nucleotide sequence encoding a reporter gene product, and wherein expression of the reporter gene product is dependent upon the presence of the negative-strand RNA virus; and
 - 10 one or more nucleotide sequences encoding each of the nucleocapsid proteins of the negative-strand RNA virus which are necessary and sufficient for replication of minigenome RNA or miniantigenome RNA synthesized by the DNA-dependent RNA polymerase.
2. The genetically engineered cell of claim 1, wherein the DNA-dependent RNA polymerase is T7 RNA polymerase, T3 RNA polymerase or SP6 RNA polymerase.
3. The genetically engineered cell of claim 2, wherein the polynucleotide is integrated into the nucleus and comprises a polymerase II promoter operably linked to a nucleotide sequence encoding the DNA-dependent RNA polymerase.
4. The genetically engineered cell of claim 2, wherein the polynucleotide comprises a noncytopathic positive-strand virus replicon.
5. The genetically engineered cell of claim 4, wherein the noncytopathic positive-strand virus replicon comprises a noncytopathic flavivirus replicon, a noncytopathic alphavirus replicon, a noncytopathic nodavirus replicon, a noncytopathic coronavirus replicon, or a noncytopathic astrovirus replicon.
6. The genetically engineered cell of claim 5, wherein the negative-strand RNA virus is an orthomyxovirus, a paramyxovirus, a rhabdovirus, a filovirus, a bornavirus or a bunyavirus.
7. The genetically engineered cell line of claim 6, wherein the negative-strand RNA virus is selected from the group consisting of: human parainfluenza virus type 1, human parainfluenza virus type 2, human parainfluenza virus type 3, human

parainfluenza virus type 4, human influenza virus, human respiratory syncytial virus
5 (RSV), measles virus, mumps virus, rabies virus, ebola virus and hanta virus.

8. The genetically engineered cell line of claim 7, wherein the reporter gene product is β -galactosidase, chloramphenicol acetyl transferase, luciferase, alkaline phosphatase, green fluorescent protein or β -glucuronidase.

9. The genetically engineered cell line of claim 8, wherein the negative-strand RNA virus is RSV, the DNA-dependent RNA polymerase is T7 RNA polymerase, the reporter gene product is β -galactosidase, and the nucleocapsid proteins necessary and sufficient for replication of the minigenome are the RSV proteins N, P and L which are encoded by three
5 T7 expression plasmids.

10. A method for detecting a negative-strand RNA virus in a sample, the method comprising:

- (a) providing a genetically engineered cell which comprises
 - a polynucleotide encoding a DNA-dependent RNA polymerase;
 - 5 a cDNA comprising a minigenome and a miniantigenome of the negative-strand RNA virus operably linked to a promoter for the DNA-dependent RNA polymerase, wherein the miniantigenome comprises a nucleotide sequence encoding a reporter gene product, and wherein expression of the reporter gene product is dependent upon the presence of the negative-strand RNA virus; and
- 10 one or more nucleotide sequences encoding each of the nucleocapsid proteins of the negative-strand RNA virus which are necessary and sufficient for replication of minigenome RNA or miniantigenome RNA synthesized by the DNA-dependent RNA polymerase;
- (b) culturing the cell for a time sufficient to synthesize the minigenome RNA or
15 the miniantigenome RNA and to express the nucleocapsid proteins;
- (c) incubating the cell with the sample; and
- (d) detecting expression of the reporter gene product.

11. The method of claim 10, wherein the DNA-dependent RNA polymerase is T7 RNA polymerase, T3 RNA polymerase or SP6 RNA polymerase.

12. The method of claim 11, wherein the polynucleotide is integrated into the nucleus and comprises a polymerase II promoter capable of causing transcription of the polynucleotide.

13. The method of claim 11, wherein the polynucleotide comprises a transcript of a noncytopathic positive-strand virus replicon.

catgcggtt cgccgacctc atgggataca ttccgctcgt cggcgcccc ttagggggcg 780
ctgccagggc cctggcgcat ggcgtaggg ttctggagga cggcgtaac tatgcaacag 840
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<211> 3010

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<213> Hepatitis C virus

<400> 34

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35 40 45

Thr Arg Lys Thr Ser Glu Arg Ser Gln Pro Arg Gly Arg Arg Gln Pro

50 55 60

Ile Pro Lys Ala Arg Arg Pro Glu Gly Trp Ala Trp Ala Gln Pro Gly

65 70 75 80

Tyr Pro Trp Pro Leu Tyr Gly Asn Glu Gly Leu Gly Trp Ala Gly Trp

85 90 95

Leu Leu Ser Pro Arg Gly Ser Arg Pro Ser Trp Gly Pro Ile Asp Pro

100 105 110

Arg Arg Arg Ser Arg Asn Leu Gly Lys Val Ile Asp Thr Leu Thr Cys

115 120 125

Gly Phe Ala Asp Leu Met Gly Tyr Ile Pro Leu Val Gly Ala Pro Leu

130 135 140

Gly Gly Ala Ala Arg Ala Leu Ala His Gly Val Arg Val Leu Glu Asp

145 150 155 160

Gly Val Asn Tyr Ala Thr Gly Asn Leu Pro Gly Cys Ser Phe Ser Ile

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Phe Leu Leu Ala Leu Leu Ser Cys Leu Thr Ile Pro Ala Ser Ala Tyr

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Glu Val Arg Asn Ala Ser Gly Val Tyr His Val Thr Asn Asp Cys Ser

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Asn Ser Ser Ile Val Tyr Glu Ala Ala Asp Ile Ile Met His Thr Pro
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Ser Ala Met Tyr Val Gly Asp Leu Cys Gly Ser Val Leu Leu Val Ser
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Gln Leu Phe Thr Leu Ser Pro Arg Gln His Glu Thr Val Gln Asp Cys
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Trp Gly Val Leu Ala Gly Ile Ala Tyr Tyr Ser Met Val Gly Asn Trp
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Ala Lys Val Leu Val Val Met Leu Leu Phe Ala Gly Val Asp Gly His
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Gly Ile Phe Ser Pro Gly Ala Lys Gln Lys Ile Gln Leu Ile Asn Thr
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Leu Asn Thr Gly Phe Leu Ala Ala Leu Phe Tyr Thr His Ser Phe Asn
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Ser Ser Gly Cys Leu Glu Arg Met Ala Ser Cys Arg Pro Ile Asp Lys
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Phe Asp Gln Gly Trp Gly Pro Ile Thr Tyr Asp Glu Gly Pro Asp Leu
 465 470 475 480

Asp Gln Arg Pro Tyr Cys Trp His Tyr Ala Pro Arg Ser Cys Gly Ile
 485 490 495

Val Pro Ala Ser Gln Val Cys Gly Pro Val Tyr Cys Phe Thr Pro Ser
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Pro Val Val Val Gly Thr Thr Asp Arg Thr Gly Ala Pro Thr Tyr Arg
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Trp Gly Glu Asn Glu Thr Asp Val Leu Ile Leu Asn Asn Thr Arg Pro
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Pro Gln Gly Asn Trp Phe Gly Cys Thr Trp Met Asn Ser Thr Gly Phe
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Thr Lys Thr Cys Gly Gly Pro Pro Cys Asn Ile Gly Gly Ala Gly Asn
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Thr Tyr Thr Lys Cys Gly Ser Gly Pro Trp Leu Thr Pro Arg Cys Met
 595 600 605

Val Asp Tyr Pro Tyr Arg Pro Trp His Tyr Pro Cys Thr Val Asn Phe
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Thr Ile Phe Lys Val Arg Met Tyr Val Gly Gly Val Glu His Arg Leu
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 645 650 655

Arg Asp Arg Ser Glu Leu Ser Pro Leu Leu Leu Ser Thr Thr Glu Trp
 660 665 670

Gln Ile Leu Pro Cys Ser Phe Thr Thr Leu Pro Ala Leu Ser Thr Gly
 675 680 685

Leu Ile His Leu His Gln Asn Ile Val Asp Val Gln Tyr Leu Tyr Gly
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Ile Gly Ser Val Val Val Ser Leu Val Ile Lys Trp Glu Tyr Val Leu
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Leu Ser Phe Phe Leu Leu Ala Asp Ala Arg Val Cys Ala Cys Leu Trp
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Met Met Leu Leu Ile Ala Gln Ala Glu Ala Ala Leu Glu Asn Leu Val
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Pro His Tyr Lys Val Phe Leu Ala Arg Leu Ile Trp Trp Leu Gln Tyr
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Phe Thr Thr Arg Ala Glu Ala Ile Leu His Val Trp Val Pro Pro Leu
 850 855 860

Asn Val Arg Gly Gly Arg Asp Ala Val Ile Leu Leu Thr Cys Ala Val
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His Pro Asp Leu Ile Phe Asp Ile Thr Lys Leu Leu Leu Ala Val Leu
 885 890 895

Gly Pro Leu Met Val Phe Leu Ala Gly Ile Thr Arg Val Pro Tyr Phe
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Val Arg Ala Gln Gly Leu Ile Arg Ala Cys Ala Leu Ala Arg Lys Val
 915 920 925

Ala Gly Gly His Tyr Ile Gln Met Ala Leu Met Lys Leu Ala Ala Leu
 930 935 940

Thr Gly Thr Tyr Leu Tyr Asp His Leu Thr Pro Leu Arg Asp Trp Ala
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His Ala Gly Leu Arg Asp Leu Ala Val Ala Val Glu Pro Val Val Phe
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Ser Asp Met Glu Thr Lys Ile Ile Thr Trp Gly Ala Asp Thr Ala Ala
 980 985 990

Cys Gly Asp Ile Ile Leu Gly Leu Pro Val Ser Ala Arg Arg Gly Arg
 995 1000 1005

Glu Ile Leu Leu Gly Pro Ala Asp Ser Leu Glu Gly Gln Gly Trp Arg
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Gly Val Arg Thr Ile Thr Thr Gly Ala Pro Ile Thr Tyr Ser Thr Tyr
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Gly Lys Phe Leu Ala Asp Gly Gly Cys Ser Gly Gly Ala Tyr Asp Ile
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Val Leu Ala Thr Ala Thr Pro Pro Gly Ser Val Thr Val Pro His Pro
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Gly Lys Ala Ile Pro Ile Glu Thr Ile Lys Gly Gly Arg His Leu Ile
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Phe Cys His Ser Lys Lys Lys Cys Asp Glu Leu Ala Ala Lys Leu Ser
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Val Ile Pro Thr Ser Gly Asp Val Val Val Val Ala Thr Asp Ala Leu
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Asp His Leu Glu Phe Trp Glu Ser Val Phe Thr Gly Leu Thr His Ile
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Ser Glu Ile Thr Leu Thr His Pro Val Thr Lys Tyr Ile Met Ala Cys
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Gly Val Leu Ala Ala Leu Ala Ala Tyr Cys Leu Thr Thr Gly Ser Val
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Val Ile Val Gly Arg Ile Ile Leu Ser Gly Lys Pro Ala Val Ile Pro
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Asp Arg Glu Val Leu Tyr Arg Glu Phe Asp Glu Met Glu Glu Cys Ala
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Lys Gln Lys Ala Leu Gly Leu Leu Gln Thr Ala Thr Lys Gln Ala Glu
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Ala Ala Ala Pro Val Val Glu Ser Lys Trp Arg Ala Leu Glu Ala Phe
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Trp Ala Lys His Met Trp Asn Phe Ile Ser Gly Ile Gln Tyr Leu Ala
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Gly Leu Ser Thr Leu His Gly Asn Pro Ala Ile Ala Ser Leu Met Ala
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Phe Thr Ala Ser Ile Thr Ser Pro Leu Thr Thr Gln His Thr Leu Leu
1795 1800 1805

Phe Asn Ile Leu Gly Gly Trp Val Ala Ala Gln Leu Ala Pro Pro Ser
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Ala Ala Ser Ala Phe Val Gly Ala Gly Ile Ala Gly Ala Ala Val Gly
1825 1830 1835 1840

Ser Ile Gly Leu Gly Lys Val Leu Val Asp Val Leu Ala Gly Tyr Gly
1845 1850 1855

Ala Gly Val Ala Gly Ala Leu Val Ala Phe Lys Val Met Ser Gly Glu
1860 1865 1870

Val Pro Ser Thr Glu Asp Leu Val Asn Leu Leu Pro Ala Val Leu Ser
1875 1880 1885

Pro Gly Ala Leu Val Val Gly Val Val Cys Ala Ala Ile Leu Arg Arg
1890 1895 1900

His Val Gly Pro Gly Glu Gly Ala Val Gln Trp Met Asn Arg Leu Ile
1905 1910 1915 1920

Ala Phe Ala Ser Arg Gly Asn His Val Ser Pro Thr His Tyr Val Pro
1925 1930 1935

Glu Ser Asp Ala Ala Ala Arg Val Thr Gln Ile Leu Ser Ser Leu Thr
1940 1945 1950

Val Thr Gln Leu Leu Arg Arg Leu His Gln Trp Ile Asn Glu Asp Cys
1955 1960 1965

Ser Thr Pro Cys Ser Gly Ser Trp Leu Arg Asp Val Trp Asp Trp Ile
1970 1975 1980

Cys Thr Val Leu Thr Asp Phe Lys Thr Trp Leu Gln Ser Lys Leu Leu
1985 1990 1995 2000

Pro Arg Leu Pro Gly Val Pro Phe Phe Ser Cys Gln Arg Gly Tyr Arg
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Ser Pro Gly Glu Ile Asn Arg Val Ala Ser Cys Leu Arg Lys Leu Gly
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Val Pro Pro Leu Arg Ala Trp Arg His Arg Ala Arg Ser Val Arg Ala
 2915 2920 2925

Lys Leu Leu Ser Gln Gly Gly Arg Ala Ala Thr Cys Gly Lys Tyr Leu
 2930 2935 2940

Phe Asn Trp Ala Val Lys Thr Lys Leu Lys Leu Thr Pro Ile Pro Ala
 2945 2950 2955 2960

Ala Ser Gln Leu Asp Leu Ser Gly Trp Phe Val Ala Gly Tyr Gly Gly
 2965 2970 2975

Gly Asp Ile Tyr His Ser Leu Ser Arg Ala Arg Pro Arg Trp Phe Met
 2980 2985 2990

Leu Cys Leu Pro Leu Leu Ser Val Gly Val Gly Ile Asn Leu Leu Pro
 2995 3000 3005

Asn Arg
 3010

(19) World Intellectual Property Organization
International Bureau



(43) International Publication Date
31 January 2002 (31.01.2002)

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WO 02/008447 A3

- (51) International Patent Classification⁷: C12Q 1/70 (74) Agent: G. E. EHRLICH (1995) LTD.; 28 Bezalel Street, 52 521 Ramat Gan (IL).
- (21) International Application Number: PCT/IL01/00669 (81) Designated States (*national*): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW.
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- (72) Inventors; and
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- Published:**
— with international search report
— before the expiration of the time limit for amending the claims and to be republished in the event of receipt of amendments
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- For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.*

(54) Title: NUCLEIC ACIDS AND METHODS FOR DETECTING VIRAL INFECTION, UNCOVERING ANTI-VIRAL DRUG CANDIDATES AND DETERMINING DRUG RESISTANCE OF VIRAL ISOLATES

(57) Abstract: A nucleic acid construct is provided. The nucleic acid construct includes (a) an expression cassette including: (i) a first polynucleotide region including a 5' NCR sequence of an RNA virus and at least an N-terminal portion of a core sequence of the RNA virus; (ii) a second polynucleotide region including a 3' UTR sequence of the RNA virus and at least a C-terminal portion of a polymerase sequence of the virus; and (iii) a third polynucleotide region encoding a reporter molecule, the third polynucleotide region being flanked by the first and the second polynucleotide regions; and (b) a promoter sequence being operatively linked to the expression cassette in a manner so as to enable a transcription of a minus strand RNA molecule from the expression cassette.

WO 02/008447 A3

INTERNATIONAL SEARCH REPORT

International application No.

PCT/IL01/00669

A. CLASSIFICATION OF SUBJECT MATTER

IPC(7) : C12Q 1/70

US CL : 536/ 23.1; 435/ 5, 6, 69.1, 173.3, 235.1, 320.1

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 536/ 23.1; 435/ 5, 6, 69.1, 173.3, 235.1, 320.1

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

Please See Continuation Sheet

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	Lohmann, V. Replication of Subgenomic Hepatitis C Virus in a Hepatoma Cell Line. Science 1999 Vol. 285, pages 110- 113, see Figure 1.	1, 4- 7- 15, 18- 20, 23- 25, 28, 33, and 36- 41
Y	VARNAVSKI, A.N., Stable High-Level Expression of Heterologous Genes IN Vitro and In Vivo by Noncytopathic DNA-Based Kunjin Virus Replication System Journal Of Virology May 2000, Vol 74, No. 9, pages 4394- 4403, see Figures 1, and 2 and Discussion.	1, 4- 7- 15, 18- 20, 23- 25, 28, 33, and 36- 41
Y	KHROMYKH, A.A. et al. cis- and trans- Acting Elements in Flavivirus RNA Replication Journal of Virology April 200 Vol 74, No. 7, pages 3253- 3263, see Figures 1 and 6, Table 1, and Discussion.	1, 4- 7- 15, 18- 20, 23- 25, 28, 33, and 36- 41
A	Ikeda, M. et al. Selectable Subgenomic and Genome-Length Dicistronic RNAs Derived from an Infectious Molecular Clone of the HCV-N Strain of Hepatitis C Virus Replicates Efficiently in Cultured Huh7 Cells.Journal of Virology 2002 Vol. 76, pages 2997-3006, Figure 1A, 6	1, 4- 7- 15, 18- 20, 23- 25, 28, 33, and 36- 41

☐ Further documents are listed in the continuation of Box C.☐ See patent family annex.

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document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

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Date of the actual completion of the international search

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Myron G. Hill

Telephone No. 703-308-0196

INTERNATIONAL SEARCH REPORT

International application No.

PCT/IL01/00669

Box I Observations where certain claims were found unsearchable (Continuation of Item 1 of first sheet)

This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claim Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
2. ☒ Claim Nos.: 2,3,16,17,21,22,26,27,34,35,42 and 43
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
The claims encompassed sequences not searchable because no CRF. See ERROR REPORT being sent along with this report.
3. ☐ Claim Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of Item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

☐
☐

The additional search fees were accompanied by the applicant's protest.

No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

PCT/IL01/00669

Continuation of B. FIELDS SEARCHED Item 3:

STN- MEDLINE, WEST- USPAT, PG Pub, and EPO, JPO, and Derwent
virus, replicon, reporter, infectious clone, drug, detecting, HCV, picornavirus, flavivirus

14. The method of claim 13, wherein the noncytopathic positive-strand virus replicon comprises a noncytopathic flavivirus replicon, a noncytopathic alphavirus replicon, a noncytopathic nodavirus replicon, a noncytopathic coronavirus replicon, or a noncytopathic astrovirus replicon.

15. The method of claim 14, wherein the negative-strand RNA virus is an orthomyxovirus, a paramyxovirus, a rhabdovirus, a filovirus, a bornavirus or a bunyavirus.

16. The method of claim 15, wherein the negative-strand RNA virus is selected from the group consisting of: human parainfluenza virus type 1, human parainfluenza virus type 2, human parainfluenza virus type 3, human parainfluenza virus type 4, human influenza virus, human respiratory syncytial virus (RSV), measles virus, mumps virus, rabies virus, ebola virus and hanta virus.

17. The method of claim 16, wherein the reporter gene product is β -galactosidase, chloramphenicol acetyl transferase, luciferase, alkaline phosphatase, or green fluorescent protein.

18. The method of claim 17, wherein the negative-strand RNA virus is RSV, the DNA-dependent RNA polymerase is T7 RNA polymerase, the reporter gene product is β -galactosidase, and the nucleocapsid proteins necessary and sufficient for replication of the minigenome are the RSV proteins N, P and L which are encoded by three T7 expression plasmids.

19. A kit for detecting a negative-strand RNA virus sample in a sample, the kit comprising:

a supply of genetically engineered cells which comprise:

a polynucleotide encoding a DNA-dependent RNA polymerase;

5 a cDNA comprising a minigenome and a miniantigenome of the negative-strand RNA virus operably linked to a promoter for the DNA-dependent RNA polymerase, wherein the miniantigenome comprises a nucleotide sequence encoding a reporter gene product, and wherein expression of the reporter gene product is dependent upon the presence of the negative-strand RNA virus; and

10 one or more nucleotide sequences encoding each of the nucleocapsid proteins of the negative-strand RNA virus which are necessary and sufficient for replication of minigenome RNA or miniantigenome RNA synthesized by the DNA-dependent RNA polymerase.

20. The kit of claim 19, further comprising a supply of reagents necessary to detect expression of the reporter gene product.

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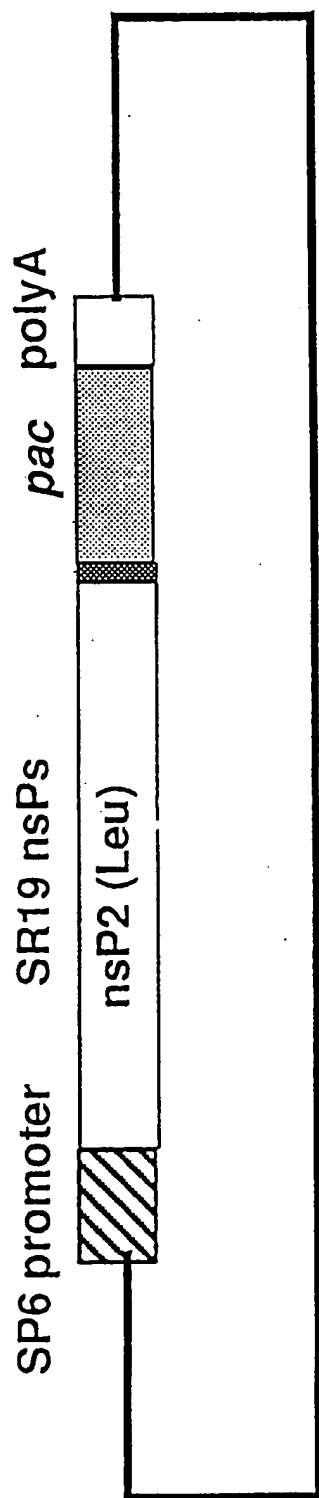


FIGURE 1A

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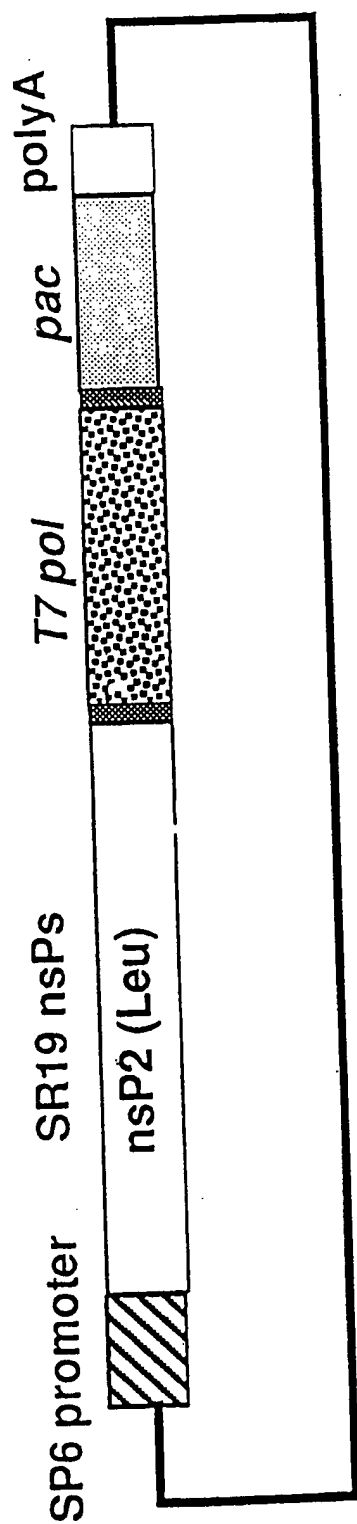


FIGURE 1B

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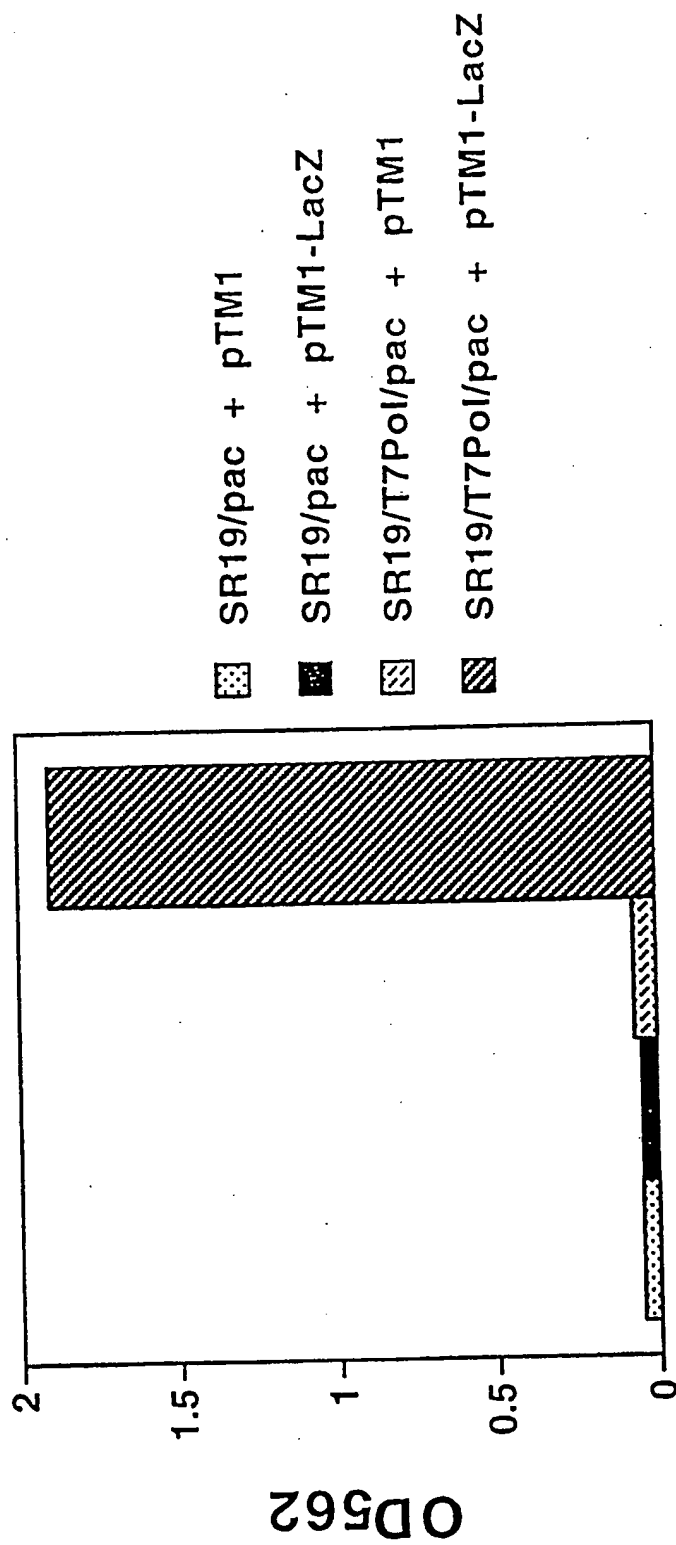


FIGURE 2

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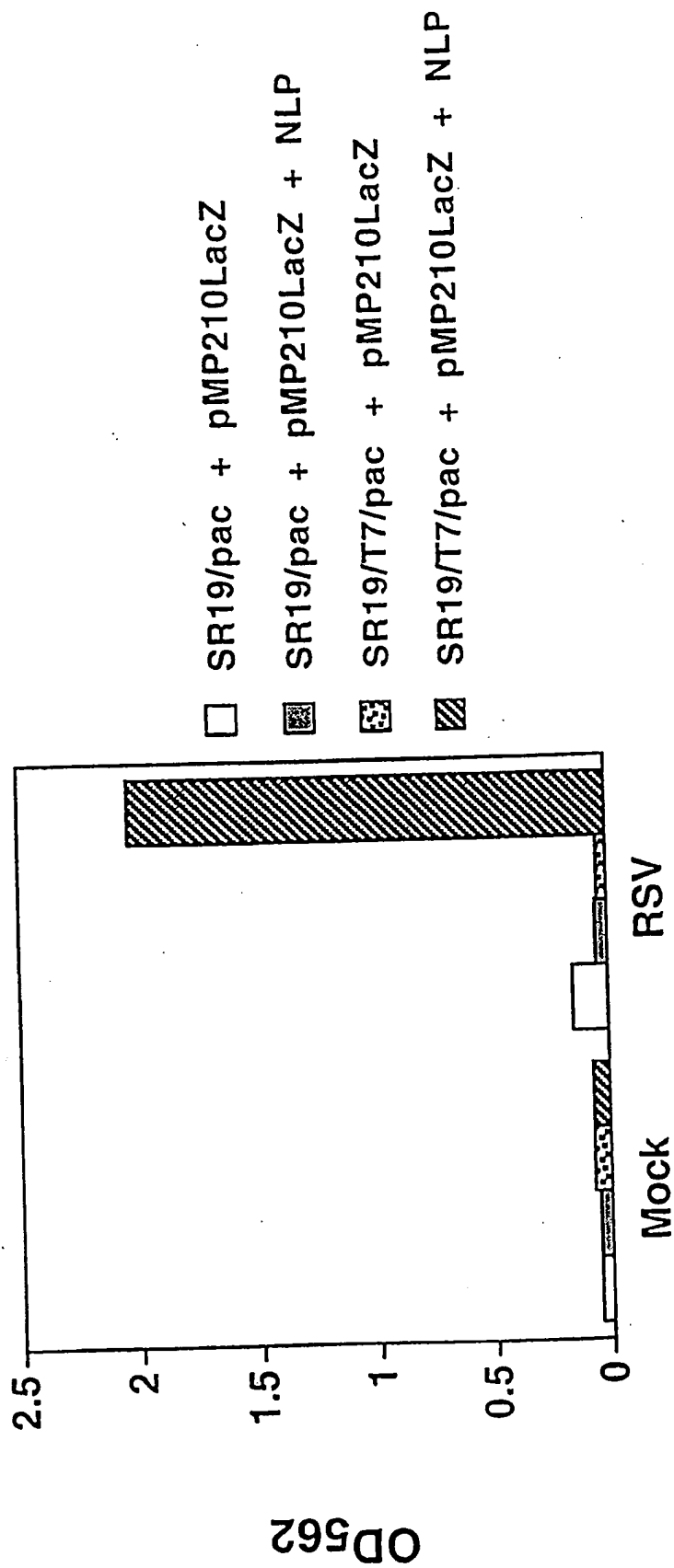


FIGURE 3A

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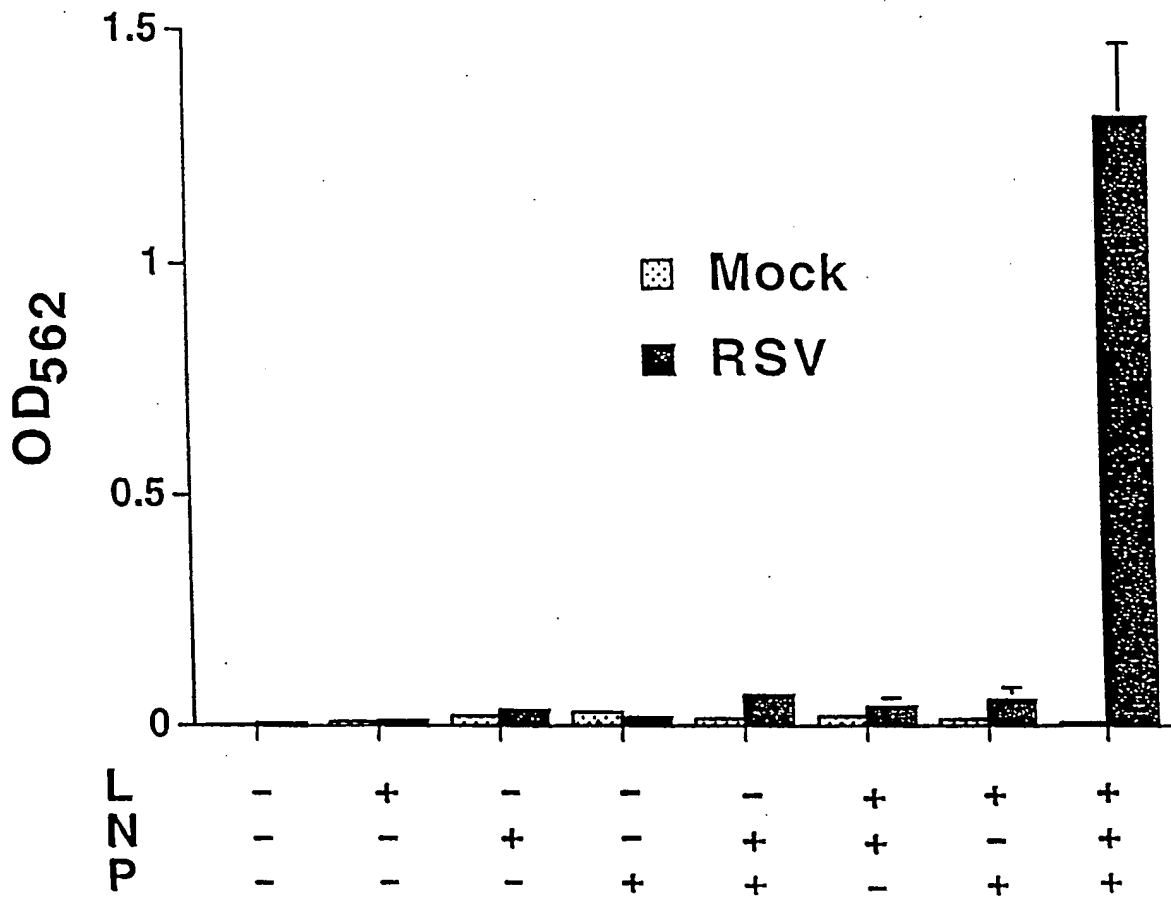


FIGURE 3B

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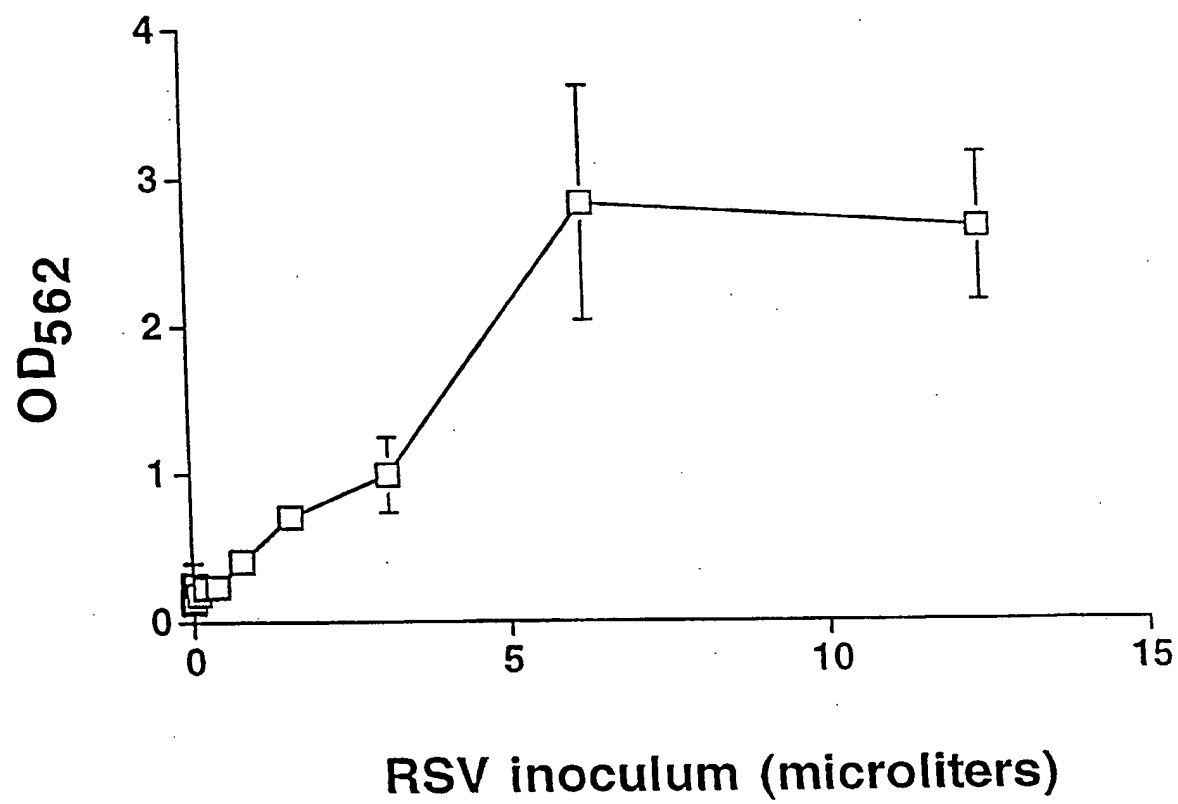


FIGURE 4A

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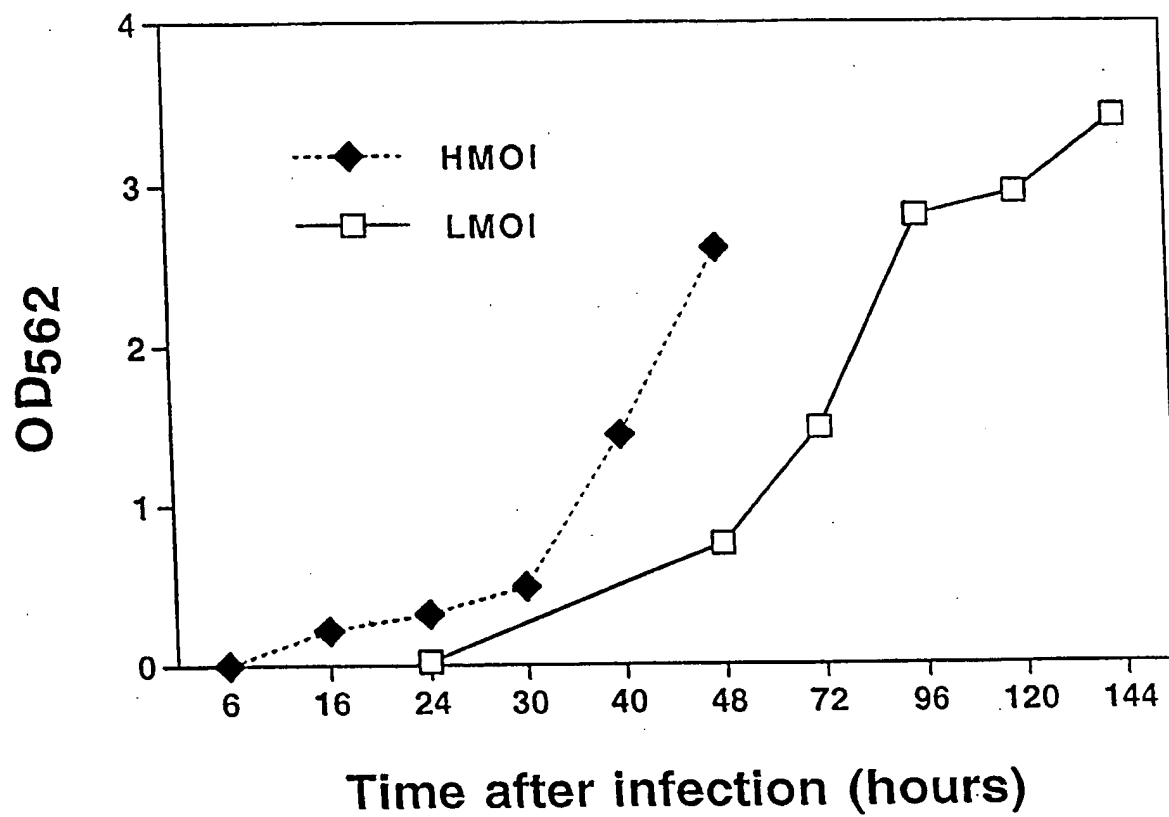


FIGURE 4B

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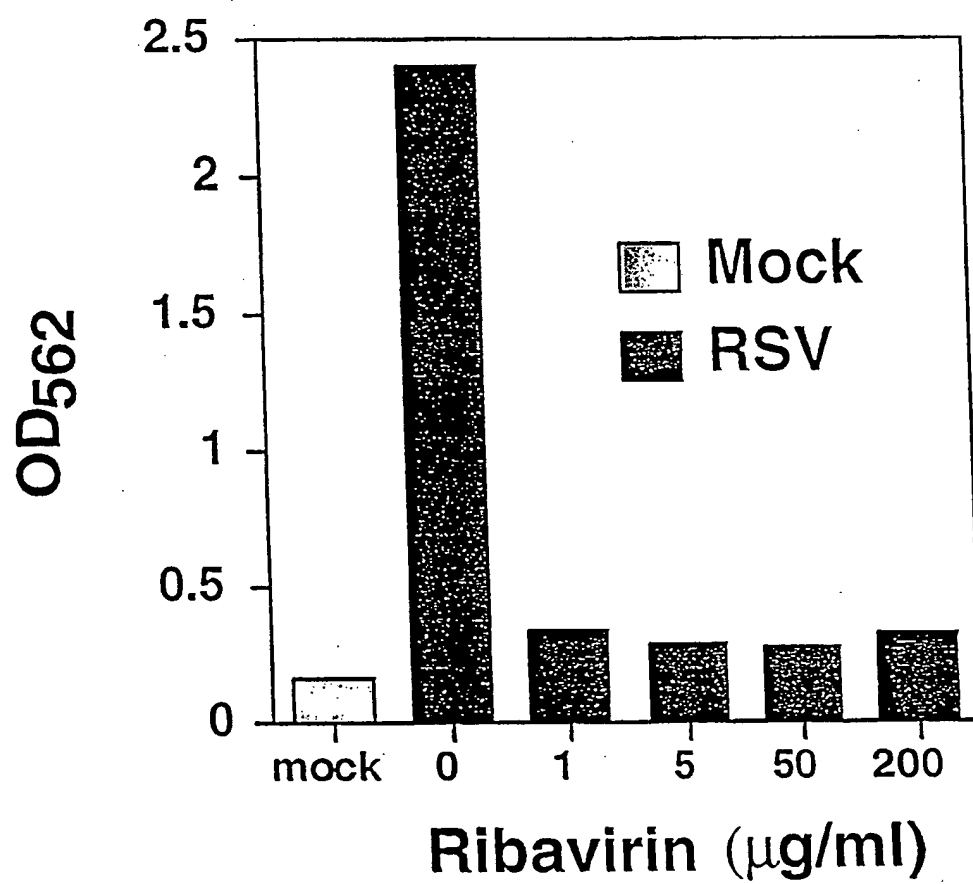


FIGURE 5A

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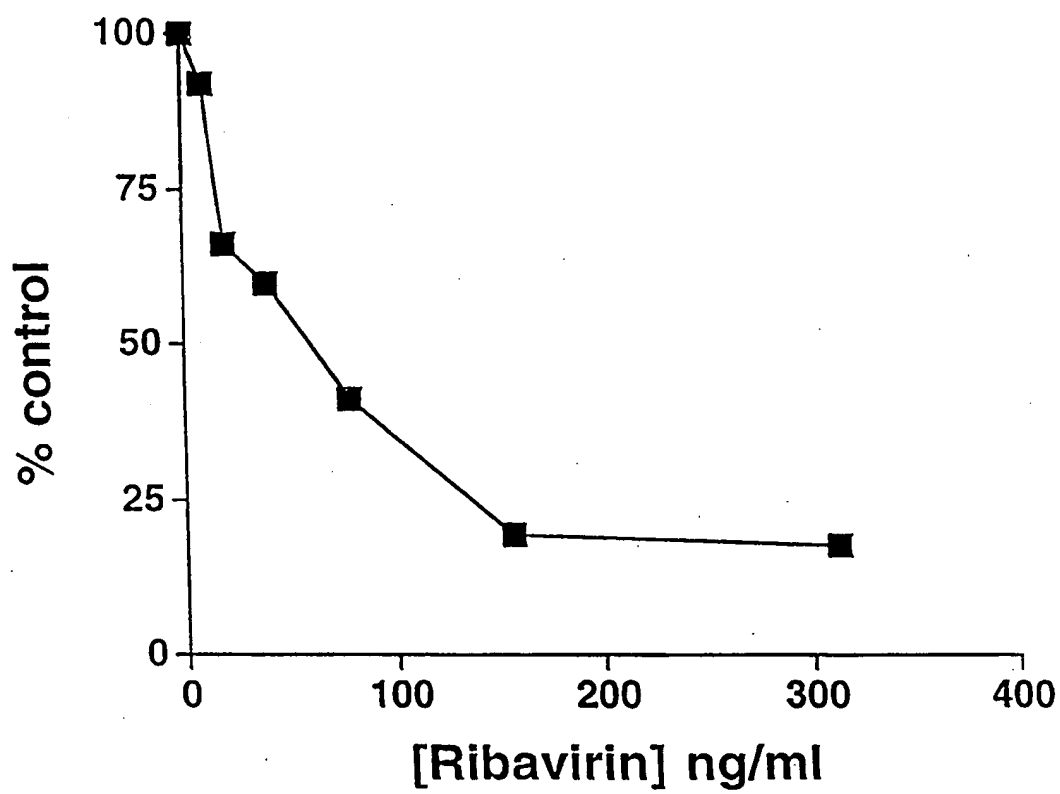


FIGURE 5B

10/11

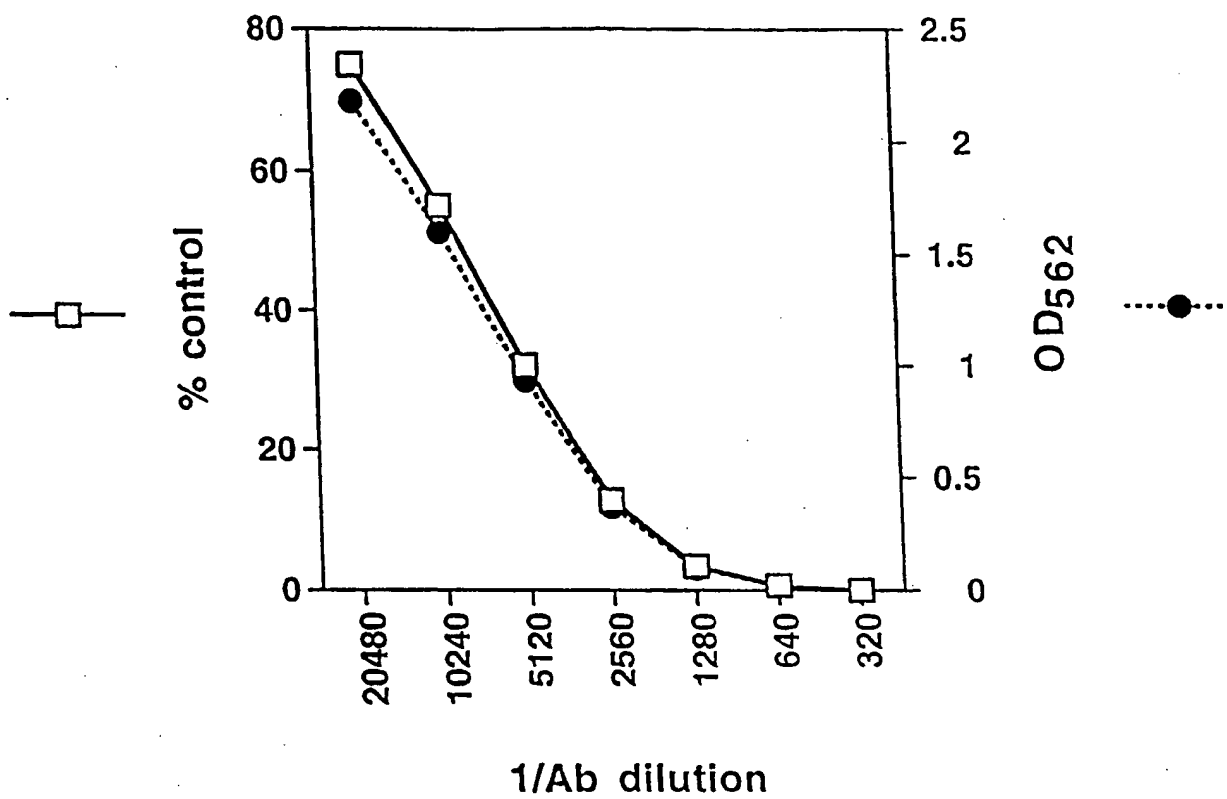


FIGURE 6

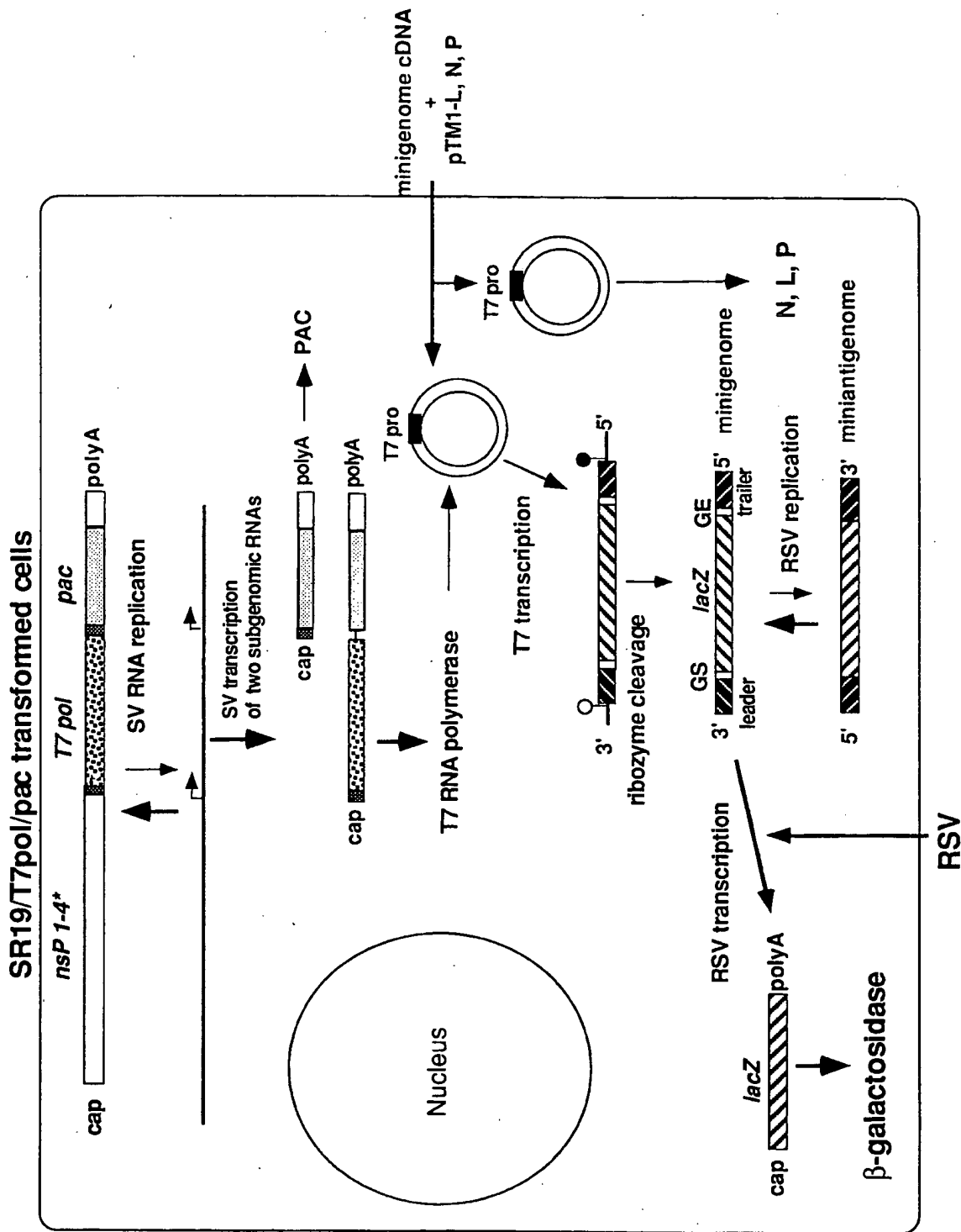


FIGURE 7

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US99/25390

A. CLASSIFICATION OF SUBJECT MATTER

IPC(7) :C12Q 1/70; C12N 5/10

US CL :435/5, 325

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 435/5, 325, 975

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

Please See Extra Sheet.

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X,P	OLIOVO et al. Detection and Quantitation of Human Respiratory Syncytial Virus (RSV) Using Minigenome cDNA and a Sindbis Virus Replicon: A Prototype Assay for Negative-Strand RNA Viruses. Virology. 10 November 1998, Vol. 251, No. 1, pages 198-205, see entire document.	1-20
A	OLIVO, P.D. et al. Transgenic Cell Lines for Detection of Animal Viruses. Clinical Microbiology Reviews. July 1996, Vol. 9, No. 3, pages 321-334, see entire document, especially page 331.	1-20

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*O document referring to an oral disclosure, use, exhibition or other means	
*P document published prior to the international filing date but later than the priority date claimed	

Date of the actual completion of the international search

24 JANUARY 2000

Date of mailing of the international search report

08 FEB 2000

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Facsimile No. (703) 305-3230

Authorized officer

MARY E. MOSHER, PH.D.

Telephone No. (703) 308-0196

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US99/25390

B. FIELDS SEARCHED

Electronic data bases consulted (Name of data base and where practicable terms used):

Medline, Derwent Biotechnology, West Derwent WPI, STN USPATFULL, East USPATFULL. Search terms: detect?, transgenic?, negative, nucleocapsid?, orthomyx?, paramyx?, rhabdo?, filo? borna?, bunya?, parainfluenza?, influenza, influenzavir?, rsv, measles, mumps, rabies, ebola, hanta?

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